Responses to reviewer RC1 (F. Moyano) on “Modelling soil CO\textsubscript{2} production and transport with dynamic source and diffusion terms: Testing the steady-state assumption using DETECT v1.0” by Edmund Ryan et al.

Reviewer’s general comments
The manuscript describes a modeling study with the main objective of determining the significance of non-steady states for determining and understanding soil respiration fluxes. The paper is well written, with a logical structure and clear sentences. Apart from some minor comments, We find the abstract correctly describes the study. The introduction is also complete and informative. The same is valid for the methods, which require a detailed description given the amount of equations and assumptions used. Overall, the study succeeds as posing a defined set of questions and methods that are then used to obtain the results. By making the data and model code available the authors make a valuable contribution to the community. The study is valid and provides some informative results as it is. However, the conclusions could be stronger with a slightly different focus. This considered, the below can be taken as suggestions for improvement unless a direct question or concern is stated.

Thank you for these positive comments.

RC1 General Comment #1
Generally, the study could focus more on the specific question posed, i.e. when are NSS conditions relevant? It could discuss less the scenario comparisons not related to this, which make the article longer than required, since they are affected by a number of factors that are not analysed properly. For example, some discussions on the response of Rsoil that are due to the source part of the model (SK) require a more detailed analysis of the functions used and could be left out. This includes precipitation effects not related to CO2 transport (as We comment below). On the other hand, a closer look at how concentrations change in soils, the amounts of air-filled pore-space and how much/fast CO2 is displaced upon wetting would be a nice addition.

Author’s Response: Thank you for these very helpful suggestions. We would like to keep the simulation experiments and the different scenarios, but we will amend the manuscript to better link them to the research questions. While there are many ways to create different simulation conditions (or scenarios), the scenarios that we selected were motivated by real data, from a real field site. The different scenarios lead to different soil conditions, thus allowing us to evaluate potential conditions or situations under which NSS conditions might be relevant.

Author’s specific changes in the manuscript:
1. We deleted ‘and CO2 production rates’ from page 7, line 21.
2. In response to your comment on “some discussion on the response of Rsoil due to Sk being left out”, we were unsure of all of the instances that you refer to so we have concentrated on deleting the parts related to ‘precipitation effects not related to CO2 transport’. In particular, we deleted the entire second paragraph of section 4.3 reducing the length of the manuscript by a page.
3. In response to your final point, we do already mention this in the second half of the second paragraph of section 4.2. We have added an extra sentence to make it clear about the change in CO2 concentration for the different soil texture types following a rain event.
**RC1 General Comment #2**

Since the NSS and SS models do not differ in the production or source of CO2, the only difference should be where this CO2 remains after being produced. So it would be very informative to include the storage state variable, i.e. how much CO2 is in the soil. The total (Rsoil + storage) should be equal for both models (otherwise there is a mass balance problem, as there is no other output flux for CO2). This also makes clearer that a NSS is always a temporal condition, so any difference (at daily or seasonal scales) should be explained by changes in storage.

**Author’s Response:** Thank you for this comment. A storage state variable for soil CO2 is already included in the DETECT model. After reading your comment in its entirety, we now realise that by a storage state variable, you mean total soil CO2 over the soil profile. You state the total (Rsoil + storage) should be equal for both models, but we think you meant to say that the total (Rsoil + change in storage) should be the same. We have checked that they are the same. Please see appendix S3 for details.

**Author’s specific changes in the manuscript:**
1. Appendix S3 in the supplementary information is new.
2. We have added new text in the manuscript at the bottom of page 8 in order to refer to appendix S3.

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**RC1 General Comment #3**

Because changes in CO2 storage can affect the net Rsoil, initial conditions that lead to a change in storage can affect the outcome. In that case it is better to get the model equilibrium to use as initial conditions instead of values fitted from data.

**Author’s Response:** Thank you for this comment. We essentially did as the reviewer suggested. The DETECT model was run during the growing season of 2007 when measurements of soil CO2 concentrations were available for three different depths as well as above ground CO2 concentration. The initial values for this 2007 run (i.e. the soil CO2 concentrations for all depths) were estimated by fitting a simple function (described in appendix S2 of the supplementary information) to the CO2 data from near the start of the 2007 growing season. The initial conditions used for 2008 (i.e. soil CO2 concentration for 1st April, 2008) were taken from the soil CO2 simulated from DETECT from the final day of the growing season for 2007 (30th September, 2007). In a follow-up paper (Samuels-Crow et al., in revision), we found that it only takes about 1-2 weeks to achieve an equilibrium state, so the model output after this initial time period should not be affected by the initial conditions.

**Author’s specific changes in the manuscript:**
1. The text that directly following equation 9a-c (which describes how the initial conditions were estimated) has been updated to improve clarity.
2. Appendix S2 (referred to by the text in the manuscript that describe the initial conditions) has been updated to improve clarity.

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**Reviewer’s specific comments**

Further questions and suggestions are given below as specific comments. Specific comments (Numbers are for the page and line)
RC1 Specific Comment #1 (3/47) The term moreover here does not seem to connect the two sentences. The second does not add to the previous.

Agreed. We have removed ‘moreover’ from this sentence in the abstract.

RC1 Specific Comment #2 (3/50-51) Integration time will surely also play a role, and NSS and SS differences will decrease for longer periods. Only a feedback of [CO2] on respiration or as a flux of dissolved inorganic C to groundwater (neither modeled) would result in different accumulated long-term Rsoil.

You are correct that NSS matters less for longer periods (e.g. years to decades), but over shorter periods (days-months) consideration of NSS condition is important. For example, when SS is not true this means that isotope methods (which can rely of this SS assumption) should not be used to partition soil respiration into its different components. This comment is similar to the second general comments from the second reviewer (RC2); please see the response this to see the changes we have made in the manuscript regarding this point.

RC1 Specific Comment #3 (4/4) A comparison with fossil fuels is misleading if not better clarified. Rsoil is part of the fast C cycle. Not necessarily a net addition of C.

This comparison is purely to help the reader appreciate the size of the global scale Rsoil aggregated over a year, however we appreciate how this could be misleading. Hence, we have added text to this first sentence of the introduction to ensure that it is clearer.

RC1 Specific Comment #4 (5/22) The hypothesis that the Rsoil spike after re-wetting is caused by pores filling with water and displacing CO2, is presented here, but not quite tested in the study.

This is something for a future study to address. We have included this soil CO2 transport process as a potential addition for a future version of the model (see the second paragraph of section 4.6.

RC1 Specific Comment #5 (10/15) How is Ψe(z) calculated? Is θsat(z) not the same as ϕT?

The air-entry potential is calculated from measurements and the formulae we use are taken from the literature. We have added extra references in section 2.1.1 to support these.

RC1 Specific Comment #6 (11/20) It is rather unusual to model the effects of volumetric moisture on respiration activity as an exponential function. This usually is an OK approximation only at the dry end of moisture content. Also strange is that when the θ and θant terms are 0 the function would equal 1. How does this make sense for a completely dry soil? There doesn’t seem to be any information here or in the cited studies of why this function type was chosen (other than that it uses both current and antecedent inputs). Changes in the dynamics of soil moisture induced by modifying precipitation patterns will affect Rsoil largely as a result of the shape of this function. It’s non-linear shape partly would explain why changing the frequency of precipitation with the same total amount would lead to different seasonal fluxes. The discussion of those differences should include this.

This point was raised by the other reviewer as one of the general comments. Please see our response to that which includes where we have made changes in the manuscript. In summary, θ at our field site never reached high enough values for respiration to decline. For completeness, however, we redid the control run using a respiration vs θ function that was bell shaped instead of exponential. We found that the time series of predicted soil respiration resulted in a very similar fit to the measurements.

RC1 Specific Comment #7 (13/eq.7) Here is another function that directly affects respiration activity and is strongly non-linearly related to moisture, as it includes the multiplier θ3. As with the f(θ, θant)
function, it changes Rsoil in response to changes in precipitation. This needs mentioning in the discussion.

This formula was taken from the Davidson et al. (2012) paper (mentioned above this formula in the manuscript) which used field data to test its suitability. We have thus adopted this formula here, but we appreciate that there are other options. We have thus added text in the discussion (see second paragraph of section 4.6).

RC1 Specific Comment #8 (14/4) ‘time-varying’

*Changed.*

RC1 Specific Comment #9 (15/11) The expression is not an equality so it does not say how exactly Ndt is calculated.

*Changed.*

RC1 Specific Comment #10 (15/eq.10) Would be nice to see this derived in the appendix.

There’s nothing to derive. The expressions in equation 10 are just the discretised (or finite differenced) version of equation 1. We put equation 1 in this form in order to be able to numerically solve it. The Haberman book (that we reference) gives a great explanation of this. New text has been added at the end of section 2.2 to make this clear.

RC1 Specific Comment #11 (16/9) Should actually cite the original derivation (by Cerling 1984)

*We have added this citation as well.*

RC1 Specific Comment #12 (16/eq.11) Since the only output is to the atmosphere, I’m guessing the depth terms are irrelevant and could be ignored in this model, unless the storage amount is of interest.

*I’m not sure I follow. This is the steady state solution to equation 1, so it has to involve a z term. In other words, there isn’t one single C pool but 100 different C pools, one for each depth.*

RC1 Specific Comment #13 (19/15) A reference for this procedure would be useful.

*Reference added towards the end of section 2.4.3.*

RC1 Specific Comment #14 (22/S) Parameter p probably has a strong impact on Rsoil. Uncertainties in this parameter would be informative.

*Yes, you’re right. Uncertainties are very important. For our study, we kept the parameters fixed, but when doing inverse modelling or uncertainty analysis we of course would want to assign a probability distribution to all parameters including this one. We have added this point as an additional improvement to consider to future versions of DETECT (second paragraph of section 4.6).*

RC1 Specific Comment #15 (22/10) Why without Cmic and CUE?

The model fitting took place a period of time prior to the DETECT model being developed, and the formula (eqn 5) in that instance was used to estimate the soil respiration of CO2 from microbial sources. At the time, we did not have measurements of C_MIC or CUE so these were left out of that version of the submodel. We have added a sentence following the sentence in question (start of third paragraph of section 2.4.5) to give the reasons stated above.

RC1 Specific Comment #16 (24) The paper makes texture a central point of the scenarios and discussion. However, the methods section did not make at all clear how texture affects the outcomes in the model. Presumably, texture is used in the HYDRUS model, thus affecting θ. Maybe
also affecting eq. 2 (but it was not specified how). Given the discussion related to texture, this should be made clearer.

Thanks for this. We’ve added two extra sentences to the methods (end of the first paragraph of section 2.5) to make this clearer.

RC1 Specific Comment #17 (26/1-2) The first sentence here is not clear. What effects?

We’ll use a different word to make it clearer what we mean.

RC1 Specific Comment #18 (32/3-23) This paragraph almost seems too out of topic. While the model could be used to explain some of the dynamics of post-wetting Rsoil, this does not seem to be the focus of the study. As commented above, these differences induced by changes in precipitation are strongly affected by the functions using θ, which are not really analyzed here. Since the paper is rather long, it would seem preferable to leave a more careful analysis of this topic for another paper.

Thanks for this. We agree that the paper is rather long, so we have removed this second paragraph in section 4.3 as you suggested.
Responses to reviewer RC2 (anonymous) on “Modelling soil CO₂ production and transport with dynamic source and diffusion terms: Testing the steady-state assumption using DETECT v1.0” by Edmund Ryan et al.

Reviewer’s general comments
In their manuscript, Ryan et al. study under which conditions soil CO₂ production is in steady state with CO₂ fluxes at the soil surface using a modelling approach, in which they focus on the effects of grain size and antecedent temperature and soil moisture conditions. Therefore, the authors present a new model of non-steady-state soil CO₂ production (DETECT v1.0) and compare the model results with a simplified version of the model which assumes steady state conditions (no delay between subsoil production of CO₂ and CO₂ the flux at the soil surface), by applying the model to an experimental site in Wyoming (PHACE).

RC2 General Comment #1
The authors address some important questions: which environments factors control subsoil CO₂ production and how can these processes be correctly simulated using a modelling approach. Overall, the manuscript is well-written and has a good structure. The abstract is informative and provides a good overview of the questions the authors address and a brief overview of the set-up of the study. The introduction gives an overview of the studied subject and existing knowledge, although it could be shortened in my opinion (see specific comments). The methodology provides a complete overview of the structure of the DETECT model and the equations it uses. At some points, however, some information is still missing (see specific comments). In the results section, the authors present how they applied the model to assess the effect of different environmental factors supported by clear graphs. In the discussion section, in my opinion, the authors should focus more on the processes lying at the basis of their observations, such as the effect of soil moisture on microbial and root CO₂ respiration (see specific comments). The fact that the authors provide the codes of their model together with a clear user manual increases the impact of their contribution.

Author’s Response: Thank you for these comments. We will ensure we carefully and fully address the specific comments you refer to here.

Although I believe that this manuscript provides a valuable contribution to existing knowledge on how to model CO₂ production in soils, I have some concerns and suggestions, as formulated below and in the specific comments.

RC2 General Comment #2
A main concern is that most of the different amounts of modelled Rsoil between the scenarios arise from the effect that soil moisture has on the production of CO₂ from both sources (roots and microbes), e.g. as shown in Figure 2 between days 220 and 240. The effect of soil moisture on CO₂ production by both roots and microbes is regulated by equation 4a, which assumes an exponential relationship between θ and the amount of CO₂ respiration. The conclusion that precipitation regime characteristics and/or including antecedent soil moisture and temperature conditions have an impact on the magnitude of the soil CO₂ efflux (as formulated in the conclusion) is thus greatly affected by the use of eq. 4a. Using a different equation in which e.g. CO₂ respiration rates decrease at very high soil moisture contents, might thus lead to a different conclusion. E.g., using a soil moisture – respiration response function in which CO₂ production is inhibited at very high soil moisture levels might lead to less CO₂ respiration using NSS conditions. Therefore, I would encourage a more elaborate discussion (in addition to P33 L11-13) on the effect of this equation on
your results or, better, an assessment of how including a different soil moisture - respiration response function affects the model results. Moreover, it should be more clearly explained how eq. 4a and 4b affect the produced CO2 by roots and microbes, so this is more easily understandable for the reader.

**Author’s Response:** Thank you for this comment, and we completely understand your concern. We have evaluated an alternative production versus soil water content function (i.e. an alternative to equation 4a). This alternative function simulates the production of soil CO2 versus soil water content as a bell shaped curve. In other words, production increases as soil water content increases but only up until an optimum soil water content value. When soil water is higher than this value, the production decreases. The formula for this alternative function is given in appendix S4 of the revised supplementary information. A graphical representation of the original function (equation 4a) and this alternative function (appendix S4) is shown in figure S8 of the supplementary information. For the Wyoming field site, however, soil water content never reached values that would have resulted in reduced ecosystem or soil CO2 flux or respiration rates. Hence, the graphical representation of this alternative soil CO2 production function shows production increasing for values of soil water content up to the optimum soil water content value. We ran the DETECT model for March-September, 2008, using this alternative production versus soil water content function, and the time series of predicted soil respiration is given in figure S9. The predicted soil respiration fits the ecosystem respiration and microbial respiration measurements equally well, when comparing this figure with the corresponding figure in the manuscript (figure 2), which used an exponential function for the production versus soil water content relationship. Thus, either function (bell-shaped or exponential) is equally good at representing the relationship between soil CO2 production and soil water content at our well-drained, mid-latitude field site. To address your final point about how equations 4a and 4b affect the CO2 produced by roots and microbes, figure S10 in the supplementary information shows modelled S (production term in equation 1) against soil water content with the points colour coded according to three soil temperature bands.

**Author’s specific changes in the manuscript:**
1. I have added text to the end of the second paragraph of section 2.1.2 in order to briefly describe the supplementary information to applying an alternative soil CO2 production versus soil water content function (i.e. an alternative version of equation 4a which has a bell-shaped form instead of an exponential form). The details of this alternative version of equation 4a as well as accompanying figures is given in the supplementary information (Appendix S4, figure S9).
2. In response to the final point of this comment, we refer to figure S10 in the new text at the end of third paragraph of 2.1.2.

**RC2 General Comment #3**
The authors state that a correct simulation of CO2 respiration in soils can improve modelling soil C processes. Therefore it would be interesting to assess the effect of the NSS vs SS approach on the total SOC pool: does the increase in CO2 respiration using the NSS conditions lead to substantially decreasing SOC pool, or is this effect limited? Or in other words, is a correct simulation (NSS vs SS) of CO2 respiration necessary in order to correctly model changes in the total SOC pool?

**Author’s Response:** This is an interesting question, but DETECT is not designed to follow slowly changing soil carbon pools. That is, we can’t use DETECT to infer changes in the SOC pools (denoted C_SOM in model description). SOC is an input to the model (here, field data inform SOC, but other models focused on soil carbon pools could also be linked to DETECT), so the model does not predict changes to SOC. DETECT is most useful for understanding and modelling fast-time scale processes (e.g. fluxes) given known, measured, or hypothesized pool sizes (e.g. SOC). A future model development would be to couple DETECT to a dynamic soil C pool model, but this is unrealistic for the manuscript revision. Our view is that C_SOM will not differ between the SS and NSS models because
the total amount of CO2 lost from the profiles was the same over a growing season. The NSS is only important in clayey soils exposed to wetting/drying cycles and only for CO2 efflux on periods of weeks-months. The C_SOM pool would take years to change.

**Author's specific changes in the manuscript:**
1. We have included a sentence as part of second paragraph of section 4.6 which describes potential future developments of the DETECT model. In particular, we describe the possibility of coupling DETECT with a dynamic soil C pool model.

**Other suggestions and remarks are formulated in the specific comments below.**

**RC2 Specific Comment #1** (P 4 L17-18): in addition to delays due to CO2 transport times, is also something known about the effect on additional CO2 production (as this is one of the outcomes of the study)?

*We don’t understand what the reviewer means by this question. In particular, we don’t understand what ‘effect on additional CO2 production’ as an ‘outcome of the study’ means. The aim of the study is to determine if it is reasonable to assume that soil CO2 produced in the soil is respired at the same time point (steady-state), under different soil textures and precipitation regimes.*

**RC2 Specific Comment #2** (P5 L21): please clarify what you mean with ‘displacement of CO2’

*We have rephrased this sentence in the fourth paragraph of the introduction to make it clearer.*

**RC2 Specific Comment #3** (P6-7 L18-13): In my opinion, this detailed explanation of your set-up can be formulated much shorter here, as this is explained in detail in the methods section

*Thank-you for this. We have shortened the text in this second to last paragraph of the introduction.*

**RC2 Specific Comment #4** (P8 L6-16): this is mostly a repeat of the last paragraph of the introduction and can be removed.

*This first paragraph of the methods has been removed.*

**RC2 Specific Comment #5** (P8 L17 – P9 L2): If you want to shorten the manuscript I would remove this part, as this is also clear from the introduction and the rest of the methods section.

*This second paragraph of the methods has also been removed.*

**RC2 Specific Comment #6** P 11 L 20): please provide a reference for this equation

*References are already provided, but I appreciate that they might have been missed. I have moved the relevant sentence to a better place in the text that following equations 4a-c.*

**RC2 Specific Comment #7** P11 eq 3): It’s not clear to me how you obtained the value for RRbase, can this be stated explicitly?

*Thanks for highlighting this. RRbase should have been included with the list of alpha1, alpha2, etc... immediately following equation 4c. This has now been corrected. The description of how we estimated the values of the parameters is given in section 2.4.5. For RRbase, this is given in the third paragraph of section 2.4.5.*

**RC2 Specific Comment #8** (P13 L16-17): how were these different values for the constants obtained? Please provide a reference if appropriate
Section 2.4.5 explains how these parameters were estimated. However, we appreciate that this should be made clearer to the reader. We have added a sentence to the end of the second to last paragraph of section 2.1.2.

**RC2 Specific Comment #9** (P14 L13-14): please provide the value for the atmospheric CO2 concentration that was used here.

*This is already given a few lines below equation 9c.*

**RC2 Specific Comment #10** (P16 L17 – P17 L8): This paragraph belongs to the introduction, not to the materials and methods section.

*We see your point. It’s challenging to know how to fit this first paragraph of section 2.4 into the introduction, so we have decided to delete it.*

**RC2 Specific Comment #11** (P18 L8): please be more specific about the data that was created

*This last sentence of section 2.4.2 has been replaced with two sentences that hopefully are a lot clearer.*

**RC2 Specific Comment #12** (P20 L9 – 22): It would be good if you could summarize the values of these parameters in a supplementary table, this would increase the readability and reduce the amount of text.

*These weight parameters are already included in table 2. For clarity, we now refer to Table 2 in the final sentence of section 2.4.4.*

**RC2 Specific Comment #13** (P21 L5 – 10): This can be removed in my opinion, this is also explained in the caption of the table.

*Thanks for the suggestion. We have deleted this text from the first paragraph of section 2.4.5.*

**RC2 Specific Comment #14** (P21 L12 – 20: this is also explained in Appendix S1, this can be removed either in the text or in the appendix.

*We have only deleted the second half of this text as we felt it was important to define what the R*, M*, S* and f functions are since they are listed in table 1.*

**RC2 Specific Comment #15** (P22 L5 – 7: Here you state that you obtained a value for the parameter p as the ratio of Csol to Csom. However, in eq7 you state that you calculate Csol from the p parameter. This is rather confusing: is eq. 7 actually used in the model?

*Thanks for spotting this. We have measurement of C_SOL and C_SOM but only a very limited number. Equation (7) is simulating C_SOL for all 100 depths and all 732 time points. We’ll amend the text to make sure there is no confusion.*

**RC2 Specific Comment #16** (P22 L9: It is not clear how both parameters (Vbase and Km) were obtained through fitting the microbial respiration submodel to data. Please clarify. Also, why are Cmic and CUE left out?

*We’ll modify the text to make this clearer. For your second question, the model fitting took place a period of time prior to the DETECT model being developed, and the formula (eqn 5) in that instance was use to estimate the soil respiration of CO2 from microbial sources. At the time, we did not have measurements of C_MIC or CUE so these were left out of that version of the submodel.*

**RC2 Specific Comment #17** (P22 L16: please clarify how these values were adjusted.

*Okay, we’ll make this clearer for the revised version.*
RC2 Specific Comment #18 (P24 L6: I agree with the comment from reviewer 1 here: please clarify how texture affect the model outcomes.

The soil texture is an input into the HYDRUS model which was used to simulate soil water content and soil temperature for all depths and times. By varying the soil texture in HYDRUS, this resulted in different sets of soil water content and soil temperature values. We will make this clearer in the revised manuscript.

RC2 Specific Comment #19 (P24 L15: please provide the amount of precipitation in 2009 here.

Precipitation totals now given for 2008 and 2009.

RC2 Specific Comment #20 (P25 L4 – 10: In my opinion, it’s strange to already summarize the results before you have presented them, I would remove this paragraph as this is also clear from the rest of the results section.

Agreed. We did this in order to grab the attention of the reader at the start of the results section, but this short text at the start of the results section is also a repeat of the abstract. Hence, we have deleted this text.

RC2 Specific Comment #21 (P26 L7 – 8: the fact that Rsoil is larger when including the antecedent effect is likely to be a result of relationship between soil moisture and respiration (eq 4a), another formulation of this relationship could lead to a different result, see comment above.

We agree. You can think of the model with antecedent terms and the model without antecedent terms as two versions of the model. There are other versions that could be considered. A growing body of evidence (see introduction) suggests that antecedent conditions are important when simulating respiration (at different soil depths in this case). This is why we consider it here. The discussion section on antecedent conditions (section 4.4) goes into more detail about this and so hopefully addresses your comment.

RC2 Specific Comment #22 (P26 L9 – 11: You attribute the greater Rsoil to an increase in root respiration, while from Fig. 2 the increase in microbial respiration is even more significant and greatly contributes to the increase in total Rsoil. Why is this not mentioned in the text here?

This is actually a plotting mistake. The green line is supposed to represent respiration from root sources and the blue line should represent from microbial sources. We have fixed this for the revised version of the manuscript.

RC2 Specific Comment #23 (P27 L11: I don’t see how Fig. 3 shows that there is a greater root respiration.

Thanks for spotting this. An earlier version of figure 3 included the green and blue lines (like in fig.2). We have amended this part of the text towards the end of section 3.2.

RC2 Specific Comment #24 (P27 L16 – 20: This formulation is confusing: in the first sentence you state that different precipitation scenarios led to little difference between Rsoil predicted using SS and NSS, while in the second sentence you state that precipitation regime affects the magnitude of Rsoil predicted by SS and NSS. Please re-formulate this.

We’re unsure why you think this is confusing, when the layout in section 3.3 is the same as section 3.2 (for example). In section 3.2, the first paragraph talks about SS vs NSS for soil texture, while the second paragraph talks about the magnitude of Rsoil for different soil textures. Section 3.3 is the same except for the different precipitation regimes + there is only one paragraph because of less to say on SS vs NSS. We have tried our best to reformulate this text at the start of section 3.3 by
deleting most of the first sentence and merging it with the second sentence. We hope this is sufficient.

**RC2 Specific Comment #25** (P30 L6 – 8: from the data you show in the figures is seems like the difference in modelled Rsoil between SS and NSS at the timescale of a growing season is rather limited (e.g. the bars on the right side of Fig. 3), please clarify this. Also, in Fig. 3e I don’t see substantial differences between SS and NSS after day 218.

*The wording in the text was ‘greater differences’ (compared to the control scenario) not ‘substantial differences. The point we’re making here is that Rsoil under SS vs NSS is larger for clay soils (whether we assume antecedent conditions or not). We do accept that with the figures being quite small, it’s hard to see the daily differences. Hence, we’ve added an extra figure to the supplementary information to make this clearer (Fig. S3a, S3b).*

**RC2 Specific Comment #26** (P31 L1-4: I think this conclusion should be formulated less strong: the ‘erroneous conclusions’ depend on what you are modelling. Your results appear to show that using SS or NSS conditions does not have a large effect on e.g. the total amount of Rsoil over a whole growing season. However, if someone want to obtain detailed daily estimates of Rsoil on a (sub-) daily timescale, this is indeed important. I suggest the authors re-formulate these sentences.

*Thank-you for this comment. We’ve added extra text to this last paragraph of section 4.2, so hopefully the statement is more explicit.*

**Technical comments**

*Thank-you for spotting all of these. We have fixed all of these issues that you mentioned and highlighted the relevant text in yellow.*

P2 L34: ... down to 1 m

P3 L51: ... precipitation inputs. The DETECT model...

P5 L8: ... coarse-grained

P5 L9: fast CO2 diffusion rates

P5 L11: ... we expect coarse-grained soils

P5 L13: ... air-filled pore space

P6 L14: ... depth-invariant CO2 production rates

P7 L 16: behavior and to (no comma)

P11 L12: remove the comma before ‘and’

P18 L5: ... to 1 m depth

P20 L10: change to ‘(J previous time periods)’

P21 L20: if the SOC data you talk about is the same as shown in figure S4, you can refer to that figure here.

P23 L18:... 2013). These data were...
P30 L17: You could change this to: ... it may take about 15 minutes for a...

Figures and tables

**Figure 1 Caption:** everything after ‘... , and temporally varying bulk CO2 fluxes.’ is redundant here. You could alternatively refer to the material and methods section where this is also explained.

*We’ve deleted this text in the figure 1 caption.*

**Figure 2** - Legend: add that root and microbial contributions are simulated using the DETECT model. For easier comparison of the Rsol between the two scenarios, you could indicate the Rsol values shown in (a) on the bars in (b) - Caption: ‘see Table 2’ should be Table 3 (also in Fig. 3, 4, 6, S1 and S2)

*We’ve modified the text in the caption to address your first point, so hopefully the caption is clearer now. We didn’t understand what you meant by your second comment and so have ignored this.*

**Figure 3** - Names of the scenarios in the sub-figures could be replaced with more intuitive names, followed by the scenario name between brackets, to increase readability. - Include a legend for the grey and red lines.

*We chose the names of the scenarios after a lot of consideration. We wanted something that was short but easy to refer to (at least after consultation with table 3). We have, however, included a fuller version of the scenario names in the titles of each subplot.*

**Figure 5** - Subplots (a) and (b): as you want to make the comparison between measurements and model results, you could choose only to show the timespan for which measurements are available (and show the entire timespan in the supplement) - Legend: add ‘depth’: e.g. 3 cm depth

*We understand your point here, however we think that it’s better to include the whole time span here for consistency with the other figures (might confuse reader if using different time windows for different figures). We made the ‘depth’ change as you suggested.*

**Table 1** - Instead of grouping the variables by ‘Group1’, ‘Group2’, etc, it would be more intuitive to provide the names to which the groups refer in the table (e.g. Group 1 = microbial submodel parameters, etc.) - I would encourage the authors to include the references from where the parameter values were obtained in the table (where appropriate), now this is only described in the text.

*We have added names following group 1, etc.. so that it’s clearer for the reader. We understand that including references for each parameter would be useful here, but we are limited by space in table 1. We feel that referring to the equation where each parameter belongs is sufficient, since the reader can refer to that equation to seek the proper reference. Another reason for not including a reference for each parameter here is that some parameters are based on the references given in the text following calculations that we have described in the manuscript. Hence, it is far better for the reader to go directly to the equation where the text that follows that equation and also to section 2.4.5 (now included in the caption) to understand what each parameter is and how it was estimated.*

**Table 3** - Bottom row, middle column: ‘about’ should be ‘above’?

*Changed.*
Supplementary information

Appendix S1 - Is there any evidence that root biomass varies between 0.5 and 1.5 times the amount measured in the middle of the growing season? Please include this. - Last sentence of first paragraph: ‘decays’ should be ‘declines’?

*We’ve updated the text in appendix S1 to address this. ‘Decays’ has now changed to ‘declines’.*

Figure S2 - Same remarks as for Fig. 3

*Titles for each subplot have been updated.*
Modelling soil CO$_2$ production and transport with dynamic source and diffusion terms:

Testing the steady-state assumption using DETECT v1.0

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Abstract

The flux of \( \text{CO}_2 \) from the soil to the atmosphere (soil respiration, \( R_{\text{soil}} \)) is a major component of the global carbon cycle. Methods to measure and model \( R_{\text{soil}} \), or partition it into different components, often rely on the assumption that soil \( \text{CO}_2 \) concentrations and fluxes are in steady state, implying that \( R_{\text{soil}} \) is equal to the rate at which \( \text{CO}_2 \) is produced by soil microbial and root respiration. Recent research, however, questions the validity of this assumption. Thus, the aim of this work was two-fold: (1) to describe a non-steady state (NSS) soil \( \text{CO}_2 \) transport and production model, DETECT, and (2) to use this model to evaluate the environmental conditions under which \( R_{\text{soil}} \) and \( \text{CO}_2 \) production are likely in NSS. The backbone of DETECT is a non-homogeneous, partial differential equation (PDE) that describes production and transport of soil \( \text{CO}_2 \), which we solve numerically at fine spatial and temporal resolution (e.g., 0.01 m increments down to 1 m, every 6 hours). Production of soil \( \text{CO}_2 \) is simulated for every depth and time increment as the sum of root respiration and microbial decomposition of soil organic matter, both of which can be driven by current and antecedent soil water content and temperature, which can also vary by time and depth. We also analytically solved the ordinary differential equation (ODE) corresponding to the steady-state (SS) solution to the PDE model. We applied the DETECT NSS and SS models to the 6-month growing season period representative of a native grassland in Wyoming. Simulation experiments were conducted with both model versions to evaluate factors that could affect departure from SS: (1) varying soil texture; (2) shifting the timing or frequency of precipitation; and (3) with and without the environmental antecedent drivers. For a coarse-textured soil, \( R_{\text{soil}} \) from the SS model closely matched that of the NSS model. However, in a fine-textured (clay) soil, growing season \( R_{\text{soil}} \) was ~3\% higher under the assumption of NSS (versus SS). These differences were exaggerated in clay soil at daily time-
scales whereby $R_{\text{soil}}$ under the SS assumption deviated from NSS by up to ~20% in the 10 days following a major precipitation event. Incorporation of antecedent drivers increased the magnitude of $R_{\text{soil}}$ by 15% to 37% for coarse- and fine-textured soils, respectively. However, the responses of $R_{\text{soil}}$ to the timing of precipitation and antecedent drivers did not differ between SS and NSS assumptions. In summary, the assumption of SS conditions can be violated depending on soil type and soil moisture status, as affected by precipitation inputs. The DETECT model provides a framework for accommodating NSS conditions to better predict $R_{\text{soil}}$ and associated soil carbon cycling processes.

*Keywords*: antecedent soil water content, DETECT, diffusion model, modelling soil CO$_2$, non-steady-state, precipitation frequency, soil respiration, soil texture, steady-state.
1. Introduction

The flux of CO$_2$ to the atmosphere from the soil (i.e., soil respiration, $R_{soil}$) is one of the largest fluxes in the global C cycle, and when aggregated globally over an entire year it is approximately ten times the annual amount of CO$_2$ emitted by fossil fuel burning (Friedlingstein et al., 2014; Hashimoto et al., 2015). Moreover, global change experiments and predictions from models agree that $R_{soil}$ is expected to increase in a future climate of elevated CO$_2$ and warming (Cox, 2001; Davidson and Janssens, 2006; Piao et al., 2009; Pendall et al., 2013; Ryan et al., 2015). Therefore, monitoring $R_{soil}$ is important for quantifying and modeling the global C cycle.

Commonly, $R_{soil}$ is monitored by directly measuring surface soil CO$_2$ fluxes using various chamber methods (Luo and Zhou, 2010; Risk et al., 2011) or by estimating $R_{soil}$ from soil CO$_2$ concentrations measured at multiple depths using probe methods (Pendall et al., 2003; Tang et al., 2003; Vargas et al., 2010). The probe methods employ diffusion equations that often rely on the assumption that $R_{soil}$ at the surface is in steady state (SS) with subsurface CO$_2$ production by roots and micro-organisms (Tang et al., 2003; Lee et al., 2004; Baldocchi et al., 2006; Luo and Zhou, 2010; Vargas et al., 2010; Šimůnek et al., 2012). That is, the SS assumption essentially assumes that CO$_2$ produced by roots and microbes within the soil profile is instantaneously respired from the soil surface, effectively neglecting delays due to CO$_2$ transport times.

Partitioning $R_{soil}$ (surface flux) into its different components (e.g., sub-surface heterotrophic [microbes] versus autotrophic [root or rhizosphere] respiration) using isotope methods (Hui and Luo, 2004; Ogle and Pendall, 2015), trenching methods (Šimůnek and Suarez, 1993), or soil CO$_2$ models (Vargas et al., 2010) also relies on the SS assumption. Even simulations of the vertical movement of soil CO$_2$ through snow have employed a SS diffusion model (Monson et al. 2006).
Recent work, however, calls into question whether this SS assumption is valid most of the time or in most systems (Maggi and Riley, 2009; Nickerson and Risk, 2009).

Given the use of the SS assumption in a diverse range of settings, the aim of this study was to determine the meteorological and site specific conditions under which the SS assumption is valid, and the circumstances under which a non-steady state (NSS) model substantially improves our understanding of subsurface processes that lead to observed \( R_{\text{soil}} \). We focused on soil texture because it is a critical factor underlying soil porosity and tortuosity, which, in turn, control soil CO\(_2\) diffusion rates (Bouma and Bryla, 2000). For example, coarse-grained (e.g., high sand content) soils generally facilitate fast CO\(_2\) diffusion rates, especially under low soil moisture conditions associated with high air-filled porosity (Bouma and Bryla, 2000); the opposite is expected for finer-grained (e.g., silt or clay) soils. Thus, we expect coarse-grained soils to generally induce SS conditions for soil CO\(_2\), whereas fine-grained soils would likely produce frequent and longer duration NSS conditions, especially following rain pulses that decrease air-filled pore space, thereby reducing CO\(_2\) diffusivity.

We also focused on the impacts of precipitation variability given that the timing and magnitude of precipitation pulses can have large effects on \( R_{\text{soil}} \) (Huxman et al., 2004; Schwinning et al., 2004; Sponseller, 2007; Cable et al., 2008; Borken and Matzner, 2009; Ogle et al., 2015). Precipitation indirectly impacts \( R_{\text{soil}} \) via its influence on soil moisture dynamics, and soil moisture and soil texture affect both diffusivity (physical process) and CO\(_2\) production (primarily biological process governed by roots and microbes). For example, as precipitation pulses infiltrate the soil, the CO\(_2\) in the pore spaces gets displaced with water, which may be seen as a transient spike in \( R_{\text{soil}} \) (e.g., Lee et al., 2004). Such transient spikes, however, may also be attributable to changes in decomposition, microbial growth, and/or C substrate availability in...
response to wetting (Birch, 1958; Borken et al., 2003; Jarvis et al., 2007; Xiang et al., 2008; Meisner et al., 2013). This transient response may be followed by a depression in $R_{soil}$ since water-filled pores will ultimately slow CO$_2$ diffusion and transport (Bouma and Bryla, 2000). These linked effects imply that precipitation pulses and their effects on soil moisture are likely to impose NSS soil CO$_2$ conditions, but the manner in which such pulses impact these processes is temporally dynamic and spatially complex, and therefore difficult to measure directly.

We evaluated the importance of soil texture and precipitation variability on SS versus NSS soil CO$_2$ behavior via a simulation-based approach. To allow for the possibility of both SS and NSS behavior, we implemented a depth- and time-varying CO$_2$ transport and production model that builds on the groundbreaking work of Fang and Moncrieff (1999), Hui and Luo (2004), Nickerson and Risk (2009), Moyes et al. (2010) and Risk et al. (2012). These processes are captured by a partial differential equation (PDE) model that is grounded in diffusion theory, and solved numerically. Some current NSS models make simplifying assumptions such as assuming depth-invariant CO$_2$ production rates (e.g., Fang and Moncrieff, 1999), or assuming that production only responds to concurrent environmental conditions (e.g., Nickerson and Risk, 2009). Such simplifications may make it difficult to evaluate physical and biological conditions leading to SS versus NSS behavior.

We addressed the aforementioned shortcomings of existing NSS models with the DETECT (DEconvolution of Temporally varying Ecosystem Carbon componenTs) model, version 1.0 (v1.0), which implemented four improvements. First, we simulated soil CO$_2$ at 100 m depth increments to ensure numerical accuracy of the solutions (Haberman, 1998). Second, we estimated the soil water content and soil temperature data for all depths and all time points using a separate model (HYDRUS; Šimůnek et al., 2005; Šimůnek et al., 2008). Third, we
simulated the production of CO\textsubscript{2} by microbial and root respiration at each depth by linking these processes to existing respiration models that are typically applied to “bulk” soil (Lloyd and Taylor, 1994; Cable et al., 2008; Davidson et al., 2012; Todd-Brown et al., 2012). Fourth, we included antecedent (past) environmental and meteorological conditions as part of the functions that predict soil CO\textsubscript{2} production, due to their importance for predicting soil and ecosystem CO\textsubscript{2} fluxes (Cable et al., 2013; Barron-Gafford et al., 2014; Ryan et al., 2015). For example, soil respiration following a rain event is generally greater if the rain event follows a dry period versus a wet period (Xu et al., 2004; Sponseller, 2007; Cable et al., 2008; Thomas et al., 2008; Cable et al., 2013). Such antecedent effects may underlie the importance of biological versus physical processes in governing the transition between SS and NSS behavior.

After describing the DETECT model, we subsequently use it to explore the effects of soil texture, precipitation pulses, and antecedent conditions on the relative importance of NSS soil CO\textsubscript{2} behavior and to identify the factors giving rise to such behavior. We simulated soil CO\textsubscript{2} concentrations, CO\textsubscript{2} production, and \(R_{\text{soil}}\) under four different soil textures and three different precipitation regimes. For each scenario, we implemented the DETECT model under the assumption that soil CO\textsubscript{2} production is affected by antecedent moisture and temperature versus the assumption that only concurrent conditions matter. Data from the Wyoming Prairie Heating and CO\textsubscript{2} Enrichment (PHACE) experiment (e.g., Pendall et al., 2013; Carrillo et al., 2014a; Ryan et al., 2015; Zelikova et al., 2015; Mueller et al., 2016) were used to parameterize the model and motivated the selection of the texture and precipitation scenarios. Under the different scenarios, we compared \(R_{\text{soil}}\) predicted from the DETECT model to that of a simpler SS model, and evaluated the relative impact of SS assumptions on inferring subsurface processes (e.g., CO\textsubscript{2} production by roots and microbes) and surface CO\textsubscript{2} fluxes (i.e., \(R_{\text{soil}}\)).
2. Methods

2.1 Description of the Non Steady State DETECT Model

The PDE that underlies the DETECT model (v1.0) accounts for time- and depth-varying CO$_2$ diffusivity and CO$_2$ production by root and microbial respiration (Fang & Moncrieff, 1999). We use a pair of PDEs, one describing the soil CO$_2$ derived from root respiration (subscripted with $R$), and the other for CO$_2$ derived from microbial respiration ($M$) such that for $K = R$ or $M$:

$$\frac{\partial c_K(z,t)}{\partial t} = \frac{\partial}{\partial z} \left( D_{gs}(z,t) \frac{\partial c_K(z,t)}{\partial z} \right) + S_K(z,t)$$

(1)

c$_K$(z,t) is CO$_2$ concentration (mg CO$_2$ m$^{-3}$), $D_{gs}(z,t)$ is the effective diffusivity of CO$_2$ through the soil (m$^2$ s$^{-1}$), and $S_K(z,t)$ is the source (or production) term (mg CO$_2$ m$^{-3}$) (Fig. 1b), all of which vary by depth $z$ (meters) and time $t$ (hours). Note that $D_{gs}$ is assumed to be the same for root- and microbial-derived CO$_2$ and is thus not indexed by $K$. In this version of the model, we assumed that CO$_2$ transport within the soil profile and over time is solely governed by gaseous diffusion, and we ignored other types of CO$_2$ transport—such as diffusion in the liquid state, convection, and bulk transport via vertical movement of water—that have been shown to have a negligible contribution (Fang and Moncrieff, 1999; Kayler et al., 2010). Total soil CO$_2$ and total CO$_2$ production are given as $c(z,t) = c_R(z,t) + c_M(z,t)$ and $S(z,t) = S_R(z,t) + S_M(z,t)$, respectively. Below we describe the two main components of the PDE model: (1) CO$_2$ diffusivity, $D_{gs}$, and (2) the production terms, $S_R(z,t)$ and $S_M(z,t)$. Finally, we note that equation 1 is the mass balance equation (see appendix S3 in the supplementary information for more information).

2.1.1 Soil CO$_2$ diffusivity sub-model

The diffusivity of CO$_2$ within the soil ($D_{gs}$) depends on soil structure and water content; we
modeled $D_{gs}$ using the Moldrup function (Sala et al., 1992; Moldrup et al., 2004). We chose this formulation because it is more accurate than other common models, such as the Millington and Quirk (2000) and Penman (1981) models (Moldrup et al., 2004). Based on Moldrup et al. (2004), $D_{gs}$ ($m^2\ s^{-1}$) is defined as:

$$D_{gs}(z,t) = D_{esp}(z,t) \left[ 2\phi_{g\text{in}}(z) + 0.04\phi_{g\text{in}}(z) \right] \left( \frac{\phi_f(z,t)}{\phi_{g\text{in}}(z)} \right)^{0.75} \times \left( \frac{T_s(z,t)}{T_0} \right), \quad (2)$$

where $D_{esp}(z,t) = D_{sp} \left( \frac{T_s(z,t)}{T_0} \right)^{0.75}$ and $D_{sp} = 1.39 \times 10^{-4} \ m^2\ s^{-1}$ is the diffusion coefficient for CO$_2$ in air at standard temperature ($T_0$, 273 K) and pressure ($P_0$, 101.325 kPa); $T_s(z,t)$ is the soil temperature (Kelvin) at depth $z$ and time $t$, and $P(t)$ is the air pressure (kPa) just above the soil surface at time $t$. The remaining terms in Eqn 2 include $\phi_f(z,t)$, the air-filled soil porosity, which is related to the total soil porosity ($\phi_T$) and volumetric soil water content ($\theta$) according to $\phi_f(z,t) = \phi_t(z) - \theta(z,t)$, and $\phi_t(z)$ is defined as $1 - BD(z)/PD$, where BD and PD are the bulk density and particle density of the soil, respectively (Davidson et al., 2006); $\phi_{g\text{in}}(z)$ is the air-filled porosity at a soil water potential ($\Psi$) of -100 cm H$_2$O (about -10 kPa); $b(z)$ is a unitless parameter that is related to the pore size distribution of the soil based on the water retention curve given by $\Psi = \Psi_e(0/\theta_{sat})^{-b}$, where $\Psi_e(z)$ is the air-entry potential – calculated from measurements (Morgan et al., 2011) – and $\theta_{sat}(z)$ is the saturated soil water content (v/v).

2.1.2 CO$_2$ source (production) terms

Soil CO$_2$ can be produced in the soil ($S$ term in Eqn. 1) by five different biological processes: (i) root growth respiration, (ii) root maintenance respiration, (iii) consumption of rhizodeposits by root-associated microorganisms and associated microbial respiration, (iv) microbial...
decomposition of newly produced plant litter that has been incorporated into the soil matrix, and (v) microbial decomposition of older soil organic matter (SOM) (Pendall et al., 2004). Due to the general lack of sufficient data and process understanding to accurately separate all five sources, the DETECT model treats CO$_2$ production as the sum of two main contributions: CO$_2$ respired by (1) roots and closely associated microorganisms (the sum of (i)-(iii)), giving $S_R(z,t)$, and (2) free-living soil microorganisms (the sum of (iv)-(v)), giving $S_M(z,t)$. Such simplification based on root and microbial sources is common in models of soil CO$_2$ transport and production (Šimůnek and Suarez, 1993; Fang and Moncrieff, 1999; Hui and Luo, 2004). Although DETECT v1.0 assumes that root and microbial respiration are independent of one another, they both depend on the same environmental data (e.g., $\theta$ and $T_s$).

CO$_2$ production by root respiration is represented as the product of three terms: (i) the mass-specific base respiration rate ($R_{Rbase}$) at a reference soil temperature of $T_s = T_{ref}$ and at average soil water and antecedent temperature conditions, (ii) root mass expressed as the amount of root carbon, $C_R(z,t)$, and (iii) functions that rescale $R_{Rbase}$ to account for the effect of soil water ($\theta$), temperature ($T_s$), and their antecedent counterparts, which are determined separately for roots and microbes. For roots, antecedent soil water and temperature are denoted as $\theta_R^{ant}$ and $T_s^{ant}$, respectively. In general, $S_R(z,t)$ is given by:

$$S_R(z,t) = R_{Rbase} \cdot C_R(z,t) \cdot f(\theta(z,t), \theta_R^{ant}(z,t)) \cdot g(T_s(z,t), T_s^{ant}(z,t)) \quad (3)$$

The functional form of $C_R(z,t)$ is informed by field data on root biomass C (see Appendix S1 for complete details). The functions $f$ and $g$ are given by:

$$f(\theta, \theta_R^{ant}) = \exp\left(\alpha_1\theta(z,t) + \alpha_2\theta_R^{ant}(z,t) + \alpha_3\theta(z,t) \cdot \theta_R^{ant}(z,t)\right) \quad (4a)$$

$$g(T_s, T_s^{ant}) = \exp\left(E_s(z,t) \left(\frac{1}{T_{ref} - T_o} - \frac{1}{T_s(z,t) - T_o}\right)\right) \quad (4b)$$
\[ E_o(z, t) = E_o^* + \alpha_4 T_{S}^{\text{ant}}(z, t) \]  

Eqn 4c

\( R_{B\text{base}}, \alpha_1, \alpha_2, \alpha_3, \alpha_4, T_o, \) and \( E_o^* \) are parameters that require numerical values (Table 1; Ryan et al. 2015), \( \theta \) and \( T_s \) are informed by field data, and \( \theta_{R\text{t}}^{\text{ant}} \) and \( T_{S\text{t}}^{\text{ant}} \) are computed from the field data (described below). The temperature scaling function, \( g \) (Eqn 4b) was motivated by Lloyd and Taylor (1994) has been successfully used to describe soil and ecosystem respiration (Luo and Zhou, 2010; Cable et al., 2013; Ryan et al., 2015). \( E_o(z, t) \) is analogous to an energy of activation term that governs the apparent temperature sensitivity of \( S_B \) (Davidson and Janssens, 2006; Cable et al., 2011; Tucker et al., 2013); we assume \( E_o \) responds to antecedent temperature, reflecting a potential thermal acclimation response (Atkin and Tjoelker, 2003; Ryan et al., 2015). \( T_o \) is also related to the apparent temperature sensitivity (Cable et al., 2011), and we assume that it is invariant with depth and time (Lloyd and Taylor, 1994; Cable et al., 2013; Barron-Gafford et al., 2014; Ryan et al., 2015). While the functional forms and choice of environmental drivers used for \( f \) and \( g \) were motivated by previous analyses (Cable et al., 2013; Barron-Gafford et al., 2014), the exact functions and parameter values were based on Ryan et al. (2015) and Cable et al. (2013). Exponential functions are also used for the moisture \( (f) \) and temperature \( (g) \) scale functions to ensure \( f > 0 \) and \( g > 0 \) (Eqn 4a). The choice of an exponential form of the functions was based on Ryan et al. (2015), with graphical forms of the functions given in figure S10 (supplementary information). However, the DETECT model is flexible enough to accommodate alternative functions for \( f \) and \( g \). For example, we ran DETECT for the control scenario using a bell-shaped function that described how soil CO\(_2\) production changes with \( \theta \) (appendix S4 and Figure S8, supplementary information) as an alternative to equation 4a. For this alternative model run, the modelled \( R_{\text{soil}} \) was very similar to the modelled \( R_{\text{soil}} \) from the results of this study (Figure S9, supplementary information).
CO₂ production by microbial respiration and SOM decomposition is represented by a modified version of the Dual Arrhenius and Michaelis-Menten (DAMM) model (Davidson et al., 2012). We exclude the O₂ term, rendering the model relevant to systems that are typically unlimited by O₂ availability, such as the semi-arid site that we focus on, but we accounted for a microbial C pool (C_MIC) and a soluble soil-C pool (C_SOL) (Todd-Brown et al., 2012) such that:

\[ S_M(z,t) = V_{\text{max}}(z,t) \frac{C_{\text{SOC}}(z,t)}{K_m + C_{\text{SOC}}(z,t)} C_{\text{MIC}}(z,t) \cdot (1 - \text{CUE}) \]  

(5)

Decomposition is assumed to be an enzymatic process that follows Michaelis-Menten kinetics, where \( V_{\text{max}} \) is the maximum potential decomposition rate, and \( K_m \) (the half-saturation constant) is the amount of substrate required for the decomposition rate to reach half of \( V_{\text{max}} \). Carbon-use efficiency (CUE) represents the proportion of total C assimilated by microbes that is allocated for microbial growth (Tucker et al., 2013). We excluded a microbial death rate term (Todd-Brown et al., 2012) because we had insufficient data on death rates, and C_MIC is only ~1% of C_SOL at our study site (Carrillo and Pendall, in review).

In contrast to the original DAMM formulation, we allowed \( S_M(z,t) \) and \( V_{\text{max}}(z,t) \) to vary by depth and time, whereas existing applications of the DAMM model are generally applied to “bulk” soil (i.e., do not vary with \( z \)). We also modeled \( V_{\text{max}} \) according to the modified energy of activation function described in Lloyd and Taylor (1994), which essentially parallels Eqns 4b-4c:

\[ V_{\text{max}}(z,t) = V_{\text{base}} \cdot f(\theta, \theta_{\text{ant}}) \cdot \exp \left( E_o(z,t) \left( \frac{1}{T_{\text{ref}} - T_o} - \frac{1}{T_S(z,t) - T_o} \right) \right) \]  

(6)

\( V_{\text{base}} \) is the ‘base’ \( V_{\text{max}} \) at a reference soil temperature of \( T_{\text{ref}} \) and at mean values of current \( \theta \) and antecedent \( \theta \) and \( T_S \) (i.e., mean values of \( \theta_{\text{ant}} \) and \( T_{\text{ant}} \)). \( E_o(z,t) \) and \( f(\theta, \theta_{\text{ant}}) \) follow the same functional forms and interpretation as described for the root respiration submodel (Eqns 3 and
4a-c), except that $\theta_R^{{ant}}$ and $T_M^{{ant}}$ are used instead of $\theta_R^{{act}}$ and $T_R^{{act}}$, respectively, and different values are specified for the parameters $a_i$, $a_2$, $a_3$, $a_4$, $T_o$, and $E_o$ to reflect microbial respiration.

The values are given in table 1, and section 2.4.5 explains how the values were estimated.

Finally, $C_{\text{SOL}}$ is modeled as a function of soil organic C content at depth $z$, $C_{\text{SOM}}(z)$, based on the fraction, $p$, of $C_{\text{SOM}}(z)$ that is soluble and the diffusivity of the substrate in liquid, $D_{\text{liq}}$ (Davidson et al., 2012). The equation for $C_{\text{SOL}}$ is given by:

$$C_{\text{SOL}}(z, t) = C_{\text{SOM}}(z) \cdot p \cdot \theta(z, t)^3 \cdot D_{\text{liq}}$$

(7)

The values of $p$ and $D_{\text{liq}}$ were taken from laboratory analysis (see § 2.4.5) and Davidson et al. (2012), respectively. We assumed that $C_{\text{SOM}}(z)$ and $C_{\text{MIC}}(z)$ (see Eqn 5) are constant over time given the relatively short simulation periods we explored here (a single growing season); but the model could be easily modified to allow for time-varying $C_{\text{SOM}}$ and $C_{\text{MIC}}$. Here, $C_{\text{SOM}}(z)$ and $C_{\text{MIC}}(z)$ are simple, empirical functions that were informed by data (see Appendix S1 for details).

Moreover, while assumption of time invariant $C_{\text{SOM}}(z)$ and $C_{\text{MIC}}(z)$ is an implicit SS assumption about biological factors affecting soil CO$_2$ dynamics, this assumption allows us to isolate the importance of NSS conditions that are primarily due to physical CO$_2$ transport characteristics.

2.1.3 Soil respiration

The efflux of CO$_2$ from the soil surface (soil respiration, $R_{\text{soil}}$) is computed as:

$$R_{\text{soil}}(t) = \frac{D_g(z=0.01, t)}{\Delta z} \left( c(z=0.01, t) - c_{\text{atm}}(t) \right)$$

(8)

$D_g(z=0.01, t)$ is the diffusivity of CO$_2$ in the soil and $c(z=0.01, t)$ is the total CO$_2$ concentration (microbial- and root-derived), respectively, at $z = 0.01$ m depth and time $t$; $c_{\text{atm}}(t)$ is the CO$_2$ concentration in the atmosphere above the soil surface; and $\Delta z$ is the depth increment that the model solves for soil CO$_2$ concentration (here, $\Delta z = 0.01$ m).
2.2 Numerical implementation of the DETECT model

The numerical solution to the NSS version of the DETECT model v1.0, as described in Eqns 1-8, requires an initial condition (IC) and two boundary conditions (BCs), which we specified as:

IC: \[ c(z, t = 0) = c_0(z) \] (9a)

Upper BC: \[ c(z = 0, t) = c_{atm}(t) \] (9b)

Lower BC: \[ \frac{\partial c(z = 1, t)}{\partial z} = 0 \] (9c)

The function \( c_0(z) \) is determined and parameterized in two stages: (1) observed soil CO\(_2\) concentration data at three depths from the start of the 2007 growing season were used to parametrize a simple function that described the change in CO\(_2\) concentration for all depths; (2) the DETECT model was run forward for the growing season of 2007, then the modelled CO\(_2\) concentrations for all depths on the final day of the 2007 growing season (September 31, 2007) was used as the initial condition for running the DETECT model for 2008. See Appendix S2 in the supplementary information for specific details. We set \( c_{atm}(t) \) equivalent to 356 ppm for all \( t \), which was the average near-surface, ambient atmospheric CO\(_2\) concentration measured at the PHACE site in the 2008 growing season. Following methods of Haberman (1998), we adopted a zero-flux lower BC (Eqn 9c) due to the lack of data at or near a depth of 1 m.

We numerically solved the non-linear PDE (Eqn. 1) by employing a forward Euler discretization with a centered difference method for the depth derivative at a depth increment of \( \Delta z = 0.01 \) m. To ensure numerical stability, we calculate model outputs at a numerical time-step of \( \Delta t = \frac{dt}{N_{dt}} \), where \( dt \) is the time step at which the predicted outputs are stored (6 hours), and \( N_{dt} \) is the number of numerical time-steps. \( N_{dt} \) is computed based on the fastest (largest) diffusion coefficient at each time step such that \( N_{dt} = \frac{dt \times \max(D_{gs})}{0.5 \times (\Delta z)^2} \), where \( \max(D_{gs}) \) is the...
maximum $D_{gs}$ across all depth increments at time $t$ (Haberman, 1998). We solved Eqn. 1 separately for both root- and microbial-derived CO$_2$ concentrations, such that for $K = R$ or $M$:

$$\frac{c_K(z, t + \Delta t) - c_K(z, t)}{\Delta t} = D_{gs}(z, t) \left( \frac{c_K(z + 2 \Delta z, t) - 2c_K(z, t) + c_K(z - \Delta z, t)}{(\Delta t)^2} \right)$$

$$+ \left( \frac{D_{gs}(z + \Delta z, t) - D_{gs}(z - \Delta z, t)}{2 \Delta z} \right) \left( \frac{c_K(z - \Delta z, t) - c_K(z + \Delta z, t)}{2 \Delta z} \right)$$

$$+ S_K(z, t)$$

We rearranged Eqn. 10 to solve for $c_K(z, t+\Delta t)$, which was iterated forward for all time-steps and depth increments; total CO$_2$ concentration at each time step and depth is calculated as $c(z, t+\Delta t) = c_R(z, t+\Delta t) + c_M(z, t+\Delta t)$. For clarity, we emphasize that equation 10 is the discretized version of equation 1, which we require in order to numerically solve equation 1 (Haberman, 1998). We programmed the DETECT model v.10 and the numerical solution method in Matlab (Mathworks, 2016).

2.3 Steady-state (SS) solution to the DETECT model

A primary goal of this work was to test if soil CO$_2$ and associated $R_{soil}$ predicted from the non-steady-state (NSS) model (DETECT) could be distinguished from that of the steady-state (SS) solution. The SS version of Eqn 1, which we refer to as the SS-DETECT model, can be solved analytically as an ordinary differential equation (ODE) by setting the $\partial cK/\partial z$ term to zero (Amundson et al., 1998). As with the NSS model, we found the SS solution to Eqn. 1 separately for root- and microbial-derived CO$_2$ concentrations, $c_R'(z, t)$ and $c_M'(z, t)$, respectively. Using the upper and lower boundary conditions described for the NSS model (Eqns 9b and 9c), the analytical SS solutions at time $t$ and depth $z$ are derived by Amundson et al. (1998) and Cerling (1984). The solution is given by:
where $K=R$ and $K=M$ refers to the soil CO$_2$ from root (R) and microbial (M) sources, respectively. $S^*_K(t)$ is the depth-averaged source term for microbial or root production (averaging over 100 0.01-m increments). The soil CO$_2$ diffusivity term, $D_{gs}(z,t)$, and upper boundary condition, $c_{atm}(t)$, are the same as previously defined (Eqns 2 and 9b, respectively; Amundson et al. (1998)).

2.4 Application of the DETECT and SS-DETECT models to the PHACE site

In this subsection, we provide an overview of the study site, including the PHACE experiment, and relevant data sources from PHACE that we used to drive the DETECT and SS-DETECT models. We also summarize how we calibrated the models in the context of the PHACE site, and we highlight data that we used to informally validate the general behavior of the models. We conclude by describing the simulation experiments that we conducted to test the effects of soil texture and precipitation variability on the importance of NSS versus SS soil CO$_2$ conditions.

2.4.1 Field site and PHACE experiment

The Prairie Heating and CO$_2$ Enrichment (PHACE) field experiment is located in south-central Wyoming (latitude 41°50’N, longitude 104°42’W, elevation = 1930 m). The site is a mixed-grass prairie with a semi-arid climate characterized by long winters (mean January temperature = -2.5 °C) and warm summers (mean July temperature = 17.5 °C), with mean annual precipitation of 384 mm (Morgan et al., 2011). The vegetation is predominantly composed of two C$_3$ grasses, western wheatgrass (*Pascopyrum smithii* (Rydb.) A. Löve) and needle-and-thread grass...
(Hesperostipa comata Trin and Rupr), and a C₄ perennial grass, blue grama (Bouteloua gracilis (H.B.K.) Lag). The soil is a fine-loamy, mixed, mesic Aridic Argiustoll, and biological crusts are not present (Bachman et al., 2010).

2.4.2 Environmental driving data

We simulated the transport and production of soil CO₂ for each 0.01 m depth increment, from the surface (0 m) to a depth of 1 m, across all 732 time steps (i.e., 4 time steps per day [every 6 hours] for 183 days from April-September). To do this, we required soil environmental data consisting of water content (θ) and temperature (Tₛ) and meteorological data including precipitation, air temperature, and air pressure. The θ and Tₛ data that were used to drive the DETECT model were created using the HYDRUS software (see § 2.4.3), calibrated against actual measurements of θ and Tₛ. For the meteorological data, actual measurements from the PHACE site were used.

The PHACE experiment involved an incomplete factorial of CO₂, warming, and irrigation (6 treatment levels total), with five replicate plots per treatment level, resulting in a total of 30 instrumented plots. One of the five plots from the control treatment—ambient CO₂, temperature (no heating), and precipitation (no supplemental irrigation)—was chosen at random and had a system installed to measure soil CO₂ concentrations continuously for three different soil depths (3, 10, and 20 cm). This plot, therefore, provided the data for driving the DETECT and SS-DETECT models. Data that we used were collected during the growing season (March-September) of 2008; θ was measured hourly at three depths (5-15, 15-25, and 35-45 cm; EnvironSMART probe, Sentek Sensor Technologies, Stepney, Australia) and we used daily averages to drive the models. Tₛ was measured hourly at two depths (3 and 10 cm) using type-T
thermocouples. Hourly precipitation (mm), air temperature (°C), relative humidity (%), and surface barometric air pressure (kPa) were recorded by an automated weather station at the site.

2.4.3 High resolution environmental data

To accommodate the 0.01 m depth increments specified for the DETECT model, we used the coarse resolution field data (above) to create finer resolution driving data. For example, temporal gap-filling of the $\theta$, $T_s$, and micrometeorological datasets was required due to gaps that occurred during a small number of days (<1%, 6%, and 2.5%, respectively) as a result of instrument failure. We used data from other nearby plots to estimate the values of the missing data, but we also used cubic spline interpolation where gaps remained. Details of these gap-filling methods can be found in Ryan et al. (2015).

We used HYDRUS-1D v4.16.0090 to simulate $\theta$ and $T_s$ in 0.01 m increments from a depth of 0.01 m to 1 m (Chou et al., 2008; Šimůnek et al., 2008; Piao et al., 2009) based on precipitation data at the site. HYDRUS simulates the movement of water by solving the Richards’ equation for water movement (Richards, 1931; Chou et al., 2008; Sitch et al., 2008) and heat transport via Fickian based advection-dispersion equations. Soil hydraulic and heat transport parameters were estimated in HYDRUS using the inverse mode to solve for parameter values based on the PHACE $\theta$ (5-10, 15-25, and 35-45 cm) and $T_s$ (3 and 10 cm) data (Simunek et al., 2005; Šimůnek et al., 2008). HYDRUS was then run in forward mode based on the tuned soil hydraulic parameters to estimate $\theta$ and $T_s$ at all 100 0.01-m depth increments at 6-hourly time intervals. For consistency, HYDRUS-derived $\theta$ and $T_s$ were used as the environmental input data to the DETECT models, even at the depths for which PHACE data were available.

2.4.4 Antecedent soil water and soil temperature conditions
We explicitly evaluated the impact of antecedent (past) $\theta$ and $T_s$ conditions on CO$_2$ production by roots and microbes, motivated by prior work that estimated the relative importance of antecedent conditions and their time-scales of influence on soil and ecosystem CO$_2$ efflux (Cable et al., 2013; Barron-Gafford et al., 2014; Ogle et al., 2015; Ryan et al., 2015). Antecedent soil water content and antecedent soil temperature—$\theta_{Kant}(z,t)$ and $T_{sant}(z,t)$, respectively, for $K = R$ (roots) and $M$ (microbes)—were computed as weighted averages of the HYDRUS-produced $\theta(z,t)$ and $T_s(z,t)$ data, respectively. These calculations were done external to the DETECT model, and the antecedent variables were supplied as driving variables to DETECT. For example, for each 0.01 m increment ($z$) and time period ($t$), antecedent soil water associated with microbial CO$_2$ production was calculated as:

$$\theta_{Mant}(z,t) = \sum_{j=1}^{J} w(j) \cdot \theta(z,t-j)$$

The $w$’s are the antecedent importance weights, which sum to 1 from $j = 1$ (previous time period) to $j=J$ (previous time periods). The weights were informed by results from an analysis of ecosystem respiration at the PHACE site (Ryan et al., 2015). For microbes, $J = 4$ days and $w = (0.75, 0.25, 0, 0)$, indicating the strong importance of $\theta$ conditions occurring yesterday ($j = 1$) (Oikawa et al., 2014). Similar equations were used to compute $\theta_{Rant}(z,t)$ and $T_{sant}(z,t)$, each with their own set of weights ($w$’s) and time-scales ($J$’s). For example, the time step and $J$ for $\theta$ differ among microbes (2 days) and roots (3 weeks); for roots, $\theta_{Rant}(z,t)$ was computed as a weighted average of past, average weekly values of $\theta$, with $j$ denoting weeks into the past, for $J = 4$ weeks, and $w = (0.2, 0.6, 0.2, 0)$, indicating a strong lag response to $\theta$ conditions occurring two weeks ago (Cable et al., 2013; Ryan et al., 2015). For antecedent soil temperature, we assumed that each of the past four days were equally important by setting the $w$ vectors to (0.25, 0.25, 0.25,
0.25), for both microbes and roots (Ryan et al., 2015). The specification of $J$ and the w’s are independent of the DETECT model formulation and can be varied by the user. For clarity we summarize these weight parameters in Table 2.

2.4.5 Overview of parameterization approach using PHACE data

In general, our aim was to specify realistic values for the parameters in the DETECT model. We did not formally “fit” the DETECT model to data, but rather, we simply determined reasonable values based on simple analyses of relevant PHACE data sets, results published for the PHACE site, or results from other relevant studies. The full list of parameters is given in Table 1, and below we describe the logic behind specifying specific values in Table 1.

The depth-distributions of root biomass C ($C_R$, Eqn 3), soil microbial biomass C ($C_{MIC}$, Eqn 5), and soil organic C ($C_{SOM}$, Eqn 7) are expressed in terms of a total C content in a 1 m deep soil column ($R^*, M^*$, and $S^*$, respectively; mg C cm$^{-2}$), multiplied by the proportion of that C that occurs at depth $z$ ($f_R(z), f_M(z)$, and $f_S(z)$, respectively). See Appendix S1 (supplementary information) for details. Regarding the data, soil organic C (Fig. S5, supplementary information) was determined by combustion of acidified, root-free soil collected from 0-5, 5-15, 15-30, 30-45, 45-75, and 75-100 cm depths, using a Costech Elemental Analyzer. Microbial biomass C was determined by the chloroform fumigation and extraction in 0.05 M K$_2$SO$_4$ (Carrillo et al., 2014b). Extracts were analysed for total C on a total organic carbon analyzer (Shimadzu TOC-VCPN; Shimadzu Scientific Instruments, Wood Dale, IL, USA) after treating with 1 M H$_3$PO$_4$ (1 µl per 10 ml of extract) to remove any carbonates. Root biomass C was estimated from ash-free root biomass and elemental analysis (Carrillo et al., 2014a; Mueller et al., 2016). The solubility parameter, $\rho$, was estimated as the ratio of $C_{SOL}$ to $C_{SOM}$ using measurements of these
two quantities which were based on unfumigated extracts obtained for microbial biomass estimations as above ($C_{\text{SOL}}$) and on total C concentration in soil ($C_{\text{SOM}}$).

The values used for the base microbial respiration rates and the half-saturation constant ($V_{\text{Base}}$ [Eqn 6] and $K_m$ [Eqn 5]; Table 1) were estimated by fitting the microbial respiration submodel, but without the $C_{\text{MIC}}$ or $C_{\text{CUE}}$ terms (Eqn 5), to microbial respiration data from the PHACE control plots (Fig. S7, supplementary information). The $C_{\text{MIC}}$ and $C_{\text{CUE}}$ terms were not included in this earlier version of $S_M$ submodel – which was used for model calibration purposes – because we did not have measurements of these two variables at the time. We estimated $V_{\text{Base}}$ and $K_m$ using a Markov Chain Monte Carlo approach, identical to the approach used in Ryan et al. (2015). In the absence of root respiration data, we assumed that base root respiration ($R_{\text{Rbase}}$ [Eqn 3]; Table 1) was proportional to the microbial base rate term (Hanson et al., 2000). The parameters denoting the effects of current soil moisture (e.g., $\alpha_1$; Eqn 4a), antecedent moisture ($\alpha_2$), and the interaction between current and antecedent moisture ($\alpha_3$) on root and microbial respiration were derived from Ryan et al. (2015), also based on an analysis of ecosystem respiration ($R_{\text{eco}}$) data from PHACE. However, we adjusted the values (Table 1) by trial and error to reflect the expectation that the effects of current soil moisture should be stronger for microbial compared to root respiration because microbes tend to respond more rapidly to precipitation pulses (Risk et al., 2008), whereas root respiration is likely to show a delayed response that depends more strongly on past moisture conditions (Cable et al., 2008; Cable et al., 2013). Of the remaining two parameters describing $S_M$ (Eqns 5-6; Table 1), the value of $C_{\text{CUE}}$ was based on results from a soil incubation study conducted at a nearby site (Tucker et al., 2013), whilst our value for $D_{\text{liq}}$ was taken from Davidson et al. (2012). Three parameters ($E_o^*$, $T_m$, and $\alpha_t$; Eqns 4a-b) were shared between the $S_R$ and $S_M$ submodels, and their values were also...
obtained from Ryan et al. (2015). Finally, the parameters used for CO$_2$ diffusivity ($b$, BD, and $\phi_{10}$; Eqn 2) were based on published, site-specific data (Morgan et al., 2011).

2.4.6 Informal model validation with soil respiration measurements

We evaluated the accuracy of the DETECT model by comparing (1) predicted $R_{soil}$ (Eqn 8) against plot-level measurements of ecosystem respiration ($R_{eco}$) (see below) and (2) predicted soil CO$_2$ concentrations, $c(z,t)$, versus observed concentrations; all observed data were from the PHACE study. Since we did not rigorously parameterize the DETECT model with PHACE data, we were simply looking for reasonable, qualitative agreement between the modelled variables and the observations (e.g., similar order of magnitude, comparable temporal trends). Observed $R_{eco}$ was measured on control plots every 2-4 weeks during the target growing season, using a canopy gas exchange chamber, and instantaneous fluxes were scaled to daily rates using a linear, empirical function (Jasoni et al., 2005; Bachman et al., 2010). We assumed that $R_{soil}$ was similar to $R_{eco}$ given that aboveground biomass was <20% of total plant biomass (Mueller et al., 2016).

Measurements of microbial respiration were obtained by applying glyphosate herbicide to small subplots in May, 2008, limiting ecosystem CO$_2$ efflux to microbial sources (Pendall et al., 2013), Non-steady state soil chambers were used to estimate the resulting surface soil fluxes every two weeks around midday (Oleson et al., 2013; Ogle et al., 2016). Soil CO$_2$ concentrations were also measured with non-dispersive infrared sensors (Vaisala GM222, Finland) installed at 3, 10, and 20 cm below the soil surface, averaged on an hourly basis (Risk et al., 2008; Vargas et al., 2011; Brennan, 2013). Observations of soil [CO$_2$] for control plots were compared against predictions of $c(z,t)$ at $z = 0.03$, 0.1, and 0.2 m and at the corresponding times.

2.5 Simulation Experiments
We evaluated the impact of three potentially important factors that could affect the frequency of
NSS (Eqns 1 and 9a-c) relative to SS (Eqn 10) conditions: (1) soil texture, (2) precipitation
patterns, and (3) importance of antecedent conditions. In the control (Ctrl) scenario, we
calculated the source terms and diffusion terms (\( S_k \) and \( D_{gs} \) in Eqns 1 and 2) based on soil
environmental (\( \theta \) and \( T_S \), soil texture (sandy clay loam: 60% sand, 20% silt, 20% clay), and
meteorological data (e.g., precipitation) measured at the PHACE site in 2008. We varied soil
texture, relative to that of the site, by varying the relative amounts of sand, silt, and clay, giving
three levels (Table 3): 80% sand, 10% silt, and 10% clay (sandy loam, scenario denoted as ST-
Sa); 20% sand, 60% silt, and 20% clay (silt loam, ST-Si); 20% sand, 20% silt, and 60% clay
(clay, ST-Cl). The control (Ctrl) scenario was also paired with the observed daily precipitation
data for 2008. We explored three additional precipitation scenarios, under the control soil
texture, by shifting the daily precipitation to occur one month earlier, or one month later, or by
using precipitation data from 2009 (scenarios P-E, P-L and P-FM, respectively; Table 3). For P-
FM, we chose 2009 because it had approximately the same total precipitation between April and
September as 2008 (340mm and 348mm for 2008 and 2009, respectively), but it fell as more
frequent events of smaller magnitudes. For each texture and precipitation scenario, HYDRUS
was used to compute the corresponding \( T_S \) and \( \theta \) at the required depth and time intervals.
Specifically, the different soil texture and precipitation regimes were used as inputs for the
HYDRUS software when generating \( T_S \) and \( \theta \) for all 100 depths and all 732 time points. Hence,
the differences in soil texture and differences in precipitation regimes were implemented by
using different input files for the HYDRUS-generated \( \theta \) and \( T_S \) data.

All above scenarios assumed that antecedent conditions are not important, which was
achieved by setting all antecedent effects parameters (\( \alpha_2 \), \( \alpha_3 \), and \( \alpha_4 \); Table 1) equal to zero. We
contrasted these scenarios against ones that included antecedent conditions (thus, computed $\theta_k^{ant}$ and $T_{crit}^{ant}$ in Eqs 3 and 6) in the calculation of soil CO$_2$ production by roots ($K=R$) and microbes ($K=M$); all such scenario names were appended with "ant" (Table 3, Fig. 1a). For each scenario summarized in Table 3, we evaluated the potential for NSS conditions by comparing the predicted $R_{soil}$ produced by the DETECT model versus the SS-DETECT model.

3. Results

3.1 Control Scenarios

Soil CO$_2$ was in steady state (SS) during most of the growing season under the control soil texture (sandy clay loam) and precipitation conditions that assumed no antecedent affects (Ctrl scenario). For example, soil respiration ($R_{soil}$) predicted by the DETECT model was approximately equal to $R_{soil}$ predicted by the SS-DETECT model during times of no or little precipitation (Fig. 2a, days < 218 or > 230). Conversely, $R_{soil}$ predicted by the SS-DETECT model was temporarily greater and more variable than that predicted by the DETECT model immediately following a large precipitation event (Fig. 2a, days 218-229). However, the total cumulative $R_{soil}$ between days 92 to 274 – hereafter ‘total growing season $R_{soil}$’ – under SS (497 g C m$^{-2}$) versus NSS (498 g C m$^{-2}$) assumptions was approximately equal (a difference of ~0.2%).

The differences between the $R_{soil}$ from DETECT and DETECT-SS using the antecedent parametrization of the source terms of the models (Ctrl-ant scenario; Fig. 2b) were generally consistent with the results from the Ctrl scenario (Fig. 2a). However, the magnitude of $R_{soil}$ predicted by both the DETECT and SS-DETECT models was up to 9 gC m$^{-2}$ day$^{-1}$ greater during days following the major rain event (i.e., during days 230-243) when antecedent conditions were considered. Moreover, the incorporation of antecedent effects led to a longer delay between the
timing of the major rain event and the maximum $R_{soil}$, which occurred ~5 days later than when
only current conditions were considered (Fig. 2a vs. 2b). As a result, total growing season $R_{soil}$
was ~15% higher under the Ctrl-ant scenario (e.g., 571 gC m$^{-2}$ under NSS assumptions, Fig. 2b)
compared to the Ctrl scenario (e.g., 498 gC m$^{-2}$ under NSS, Fig. 2a). This increase in predicted
$R_{soil}$ under the Ctrl-ant scenario for days 230-243 was primarily driven by greater root respiration
(Fig. 2a vs 2b).

3.2 Effects of soil texture

Varying soil texture resulted in the greatest difference in daily $R_{soil}$ between the DETECT and
SS-DETECT models; however, integrated over the growing season, these differences were very
small (Fig. 3a,b,c). In particular, total growing season $R_{soil}$ predicted by SS-DETECT was ~1.5%
less than predicted by DETECT for soils consisting primarily of sand and silt (ST-Sa and ST-Si
scenarios; Fig. 3a,b), but was ~3.3% less for a clay dominated soil (ST-Cl scenario; Fig. 3c red
versus grey bars). These differences in $R_{soil}$ under NSS versus SS assumptions were
approximately the same for the scenarios involving antecedent effects (Figs. 3d,e,f). Despite the
minor differences at the growing season scale, notable differences emerged at the daily scale.
For example, with the largest precipitation event of the year and the 10 days that followed (days
218-248), daily $R_{soil}$ predicted by the DETECT model was on average ~2.5% less than daily $R_{soil}$
from the SS-DETECT model for the ST-Sa and ST-Si scenarios (Fig. S1). $R_{soil}$ from DETECT
was 4% greater than DETECT-SS $R_{soil}$ for the ST-Cl scenario, but when antecedent variables
were included in the models, this difference increased to 10% (Figs. 3 and S1).

Soil texture also affected the magnitude of predicted $R_{soil}$ compared to the control
scenarios, both with and without antecedent effects (Ctrl-ant and Ctrl, respectively). In
particular, we found that total growing season $R_{soil}$, whether from the DETECT or the SS-
DETECT model, was ~30% and ~60% higher for the ST-Si and ST-Cl scenarios relative to the Ctrl scenario (Figs. 3b, 3c, 4a). The change in $R_{soil}$ was negligible, however, when the sand content was increased from 60% (Ctrl) to 80% (ST-Sa) for both models (Fig. 3a, Fig. 4a). The antecedent versions of the fine-textured scenarios (ST-Si-ant and ST-Cl-ant) resulted in ~45% and ~95% increases in total growing season $R_{soil}$, respectively, compared to the Ctrl-ant scenario (Figs. 3e, 3f, 4b). As with the Ctrl-ant scenario (§ 3.1), greater root respiration following the end of the second precipitation period between days 230 and 245, primarily drove the larger percentage increases for the SL-Si-ant and SL-Cl-ant scenarios compared to the non-antecedent versions (Fig. 4b vs Fig. 4a; Fig. 4e).

### 3.3 Effects of precipitation regimes

Although varying the timing, frequency, or magnitude of precipitation led to little difference between $R_{soil}$ as predicted by the DETECT and SS-DETECT models (Fig. S2), these precipitation regimes did affect the magnitude of $R_{soil}$ predicted by both models. For example, total growing season $R_{soil}$ predicted under the alternative precipitation scenarios was lower relative to the Ctrl scenario. This decrease was relatively small (5-10%) for the non-antecedent versions of the models (Fig. 4e), but was comparatively larger (15-22%) for the antecedent versions (Fig. 4d). This reduction appears to be driven by the amount of time over which daily $R_{soil}$ responded to the second precipitation period, which occurred around day 220, 190, and 250 in the Ctrl, P-E, and P-L scenarios, respectively. Following this precipitation event, daily $R_{soil}$ achieved values around 10 g C m$^{-2}$ day$^{-1}$ for about 20 days in the Ctrl scenario (Fig. 2a, days 220-240), but for only about five days in the P-E and P-L scenarios (Fig S2a,b, after days 190 and 250, respectively). Increasing the frequency of precipitation while retaining approximately the same annual amount (i.e., scenario P-FM) resulted in daily $R_{soil}$ being consistently less than that.
of the Ctrl scenario, which led to a reduction in total growing season $R_{soil}$ in the P-FM scenario (Fig. S2c and S2c).

3.4 Effects of antecedent responses

When antecedent soil water content and soil temperature were included in the DETECT model we found that predicted $R_{soil}$ was 15% greater for the control scenario and 29-37% greater for the fine textured soil scenarios, compared to the corresponding scenarios that did not include antecedent conditions. When the sand content was 80% or for any of the different precipitation regimes, there was a negligible difference between $R_{soil}$ predicted by the antecedent versus non-antecedent parametrizations of DETECT.

Daily $R_{soil}$ predicted by the DETECT model based on the Ctrl and Ctrl-ant scenarios agreed well with observed ecosystem respiration ($R_{eco}$), but $R_{eco}$ was slightly higher than predicted $R_{soil}$ (Fig. 2a,b), which was expected since $R_{eco} = R_{soil} +$ aboveground autotrophic respiration. For the most part, this data-model agreement was similar whether the antecedent model terms were included (Fig. 2b) or not (Fig. 2a). Unfortunately, $R_{eco}$ data were not available during the time period (days 230-250) associated with the greatest disagreement between the Ctrl and Ctrl-ant scenarios. During this period, frequent hourly measurements of soil [CO$_2$] were in better agreement with predicted soil CO$_2$ from the Ctrl-ant scenario compared to the Ctrl scenario (Figs. 5a,b, S4a,b). After day ~250, based on the DETECT model, both scenarios (Ctrl and Ctrl-ant) under-predicted the observed soil [CO$_2$] by ~ 50% (Fig. 5).

4. Discussion

The DETECT and SS-DETECT models provide a framework for evaluating the circumstances under which steady-state (SS) assumptions of soil CO$_2$ production and surface soil respiration ($R_{soil}$) are valid, and to identify the major physical (i.e., soil texture, soil moisture) and/or
biological (i.e., root and microbial respiration responses) factors that lead to non-steady-state (NSS) conditions.

4.1 Steady-state versus non-steady-state conditions

At the seasonal scale, there was reasonable agreement between total growing season $R_{\text{soil}}$ predicted under the assumption of SS versus NSS conditions, but the strength of this agreement depended on soil texture (see §4.2). At the daily scale, $R_{\text{soil}}$ predicted by the DETECT model deviated from values expected under the assumption of SS conditions for 11 days or 4% of the days during the April-September growing season (Fig 2, days 218-228). These discrepancies, attributed to NSS conditions, were generally limited to periods following large rain events. For applications that assume SS conditions, such as isotopic partitioning studies (Hui and Luo, 2004; Ogle and Pendall, 2015), the SS assumption seemed reasonable during periods of minimal or no precipitation, representative of times during which soil water content changes very little or gradually. For sites or time periods characterized by pulsed precipitation patterns, our results suggested that NSS conditions would be more likely over longer periods of time.

4.2 Effect of varying soil texture

Our results indicated that soil texture exerts the strongest control over the prevalence of NSS soil CO$_2$ conditions. For a predominantly (e.g., 60%) sandy or silty soil, soil CO$_2$ transport and efflux generally aligned with the SS assumption (Fig. 2, Fig. 3a-b). This was consistent with previous work that used SS models to predict $R_{\text{soil}}$ for similar soil types (Baldocchi et al., 2006; Vargas et al., 2010).

For very fine-texture soil dominated by clay, however, SS assumptions were far less appropriate. The larger difference relative to the $Ctrl$ scenario in $R_{\text{soil}}$ predicted under SS versus NSS conditions for fine-texture (i.e., 60% clay) soil was apparent at both the growing
season scale and the daily scale following a large precipitation event (Fig. 3, S3a, S3b). In general, the DETECT model predicted that $R_{\text{soil}}$ should be higher in clay compared to sandy soil after precipitation events, a result supported by field experiments (Cable et al., 2008), but this texture effect is muted under assumptions of SS. Moreover, recovery of $R_{\text{soil}}$ to SS rates after a large rain event took ~30 days in the clay soil (Fig. 3c, days 218 to 248) compared to ~10 days for the other coarser soil texture scenarios (Fig. 2, Fig. 3a-b, days 218 to ~230). These effects of soil texture on the prevalence of NSS conditions can be attributed to soil physical properties and their effects on air-filled porosity and CO$_2$ diffusivity. Fine textured soils have smaller pores and tend to retain water for longer (Bouma and Bryla, 2000), which has the effect of decreasing soil CO$_2$ diffusivity (Fig. 6). Thus, under moist conditions that follow a rain event, it may take about 15 minutes for a CO$_2$ molecule produced at 0.5 m to diffuse to the surface in a clay soil compared to only 1-2 minutes for a sandy soil. This means that the increase in CO$_2$ concentration near the soil’s surface will be almost immediate under a coarsely textured soil (Fig. 6a), but slightly delayed under a finely texture soil. Finally, fine-textured soils have slower infiltration rates (Hillel, 1998), delaying the exposure of more deeply distributed roots and microbes to increased moisture availability. While this effect may not directly impact the SS assumption, it would lead to greater time lags between precipitation pulses and $R_{\text{soil}}$ peaks.

These findings have important implications for studies that rely on the SS assumption to predict subsurface soil CO$_2$ production. The SS assumption may be sufficient for systems defined by coarse-textured soils, but it may lead to erroneous conclusions if applied to fine-textured soils, especially at the very short-term scale (e.g. diurnal $R_{\text{soil}}$) during times of precipitation. Our simulation experiments made the simplifying assumption that soil texture is constant with depth, but in many ecosystems, texture may vary greatly with depth (Ogle et al.,
An important next step is to extend the simulations to explore the impacts of depth-varying soil texture on SS versus NSS conditions. The DETECT model can easily accommodate such modifications; allowing soil texture to vary by depth would have a direct effect on soil water content, which is simulated outside of DETECT using HYDRUS (Chou et al., 2008; Šimůnek et al., 2008; Piao et al., 2009), that can accommodate such depth variation.

4.3 Effect of varying the timing or frequency of precipitation

Unlike soil texture, varying the timing, frequency, and magnitude of precipitation resulted in predicted $R_{soil}$ that was almost identical under SS and NSS assumptions, both at the growing season and daily time-scales (Fig. S2). We had anticipated that such changes in the precipitation regime would impact SS conditions via impacts on soil air-filled porosity and potentially by changing the covariance between soil water and soil temperature, both of which affect soil CO$_2$ diffusivity (e.g., see Eqn 2). We did not explore, however, the effect of decreasing the frequency while simultaneously increasing the magnitude of individual pulses. We hypothesize that this latter scenario could produce more exaggerated or extended NSS conditions given that large rain events would infiltrate deeper, reducing CO$_2$ diffusivity across greater soil depths, thus slowing the transport of more deeply derived CO$_2$. Increasing the number of small events, as done in the $P$-$FM$ scenario, would generally confine water inputs to shallow layers, from which CO$_2$ has shorter distances to travel to reach the surface, creating less opportunity for $R_{soil}$ to exhibit NSS behavior.

4.4 Effect of antecedent conditions

The inclusion or exclusion of antecedent soil moisture and temperature effects on CO$_2$ production rates had little to no impact on the balance between SS versus NSS behavior of $R_{soil}$. However, incorporating antecedent effects generally increased the magnitude of $R_{soil}$ as
microbial respiration was stimulated more during the initial onset of the main precipitation period when antecedent effects were considered (Fig. 2b vs Fig 2a, day 218, blue line). This is expected because the instantaneous response of microbes to a rain event is expected to be greater following a dry period compared to during a wet period (Xu et al., 2004; Sponseller, 2007; Cable et al., 2008; Thomas et al., 2008; Cable et al., 2013). These dynamics are incorporated in the antecedent version of the models when the parameter corresponding to the interaction between current and antecedent soil water content is negative (e.g., $\alpha_i$, Table 1). Secondly, root respiration was greatly enhanced following the end of this period of precipitation (Fig. 2b vs Fig. 2a, days ~230-250, green line), despite there being little precipitation after day 230 (Fig. 2b). This likely occurred because our DETECT model assumed that soil water over relatively longer time periods (past 1-2 weeks, Eqn. 12) affects current root respiration rates. This partly reflects the mechanism that roots are able to take up more soil water that has infiltrated to deeper depths (Cable et al., 2013). The microbes, however, are coupled to past conditions over comparatively short time periods (a couple days).

The importance and benefit of including antecedent terms for modelling soil respiration or ecosystem respiration has been well documented (Cable et al., 2013; Barron-Gafford et al., 2014; Ryan et al., 2015). Thus, we encourage future studies to include influences of past conditions when modelling subsurface and surface CO$_2$ fluxes. Fortunately, our simulation experiments suggest that the lagged responses of microbial and root respiration to soil moisture and temperature do not have a notable impact on the SS assumption.

4.5 Comparison of modelled soil CO$_2$ with data

The good agreement between modeled and observed soil CO$_2$ concentrations—particularly when including antecedent effects—was very encouraging because the DETECT model was not
rigorously tuned or calibrated to fit data on soil [CO$_2$] or ecosystem CO$_2$ fluxes ($R_{eco}$) (Figs. 5, S4). However, there remained discrepancies between the predicted and observed CO$_2$ fluxes, particularly after rain events. These discrepancies could be an artifact of the input data used to calculate CO$_2$ production (i.e., the source term). Some parameter values were drawn from the literature and others were estimated by fitting a non-linear regression model to data. For example, the parameters describing the current and antecedent soil water content effects ($\alpha$’s) were obtained by fitting a non-linear model to $R_{eco}$ data (Ryan et al., 2015). While measured $R_{eco}$ represents both root respiration and microbial respiration contributions, it also reflects aboveground respiration, which is not currently treated in the DETECT model. Moreover, we made further assumptions about how the $R_{eco}$ parameter estimates translate to component processes (root and microbial responses), and we relied on literature information about how microbes and roots respond to precipitation events (e.g., the timing, magnitude, and lags). Future studies could rigorously fit the DETECT model to field data, such as observations of $R_{soil}$, soil CO$_2$ concentrations, and $^{13}$C isotope fluxes. Using a Bayesian methodology to do this would allow one to incorporate multiple data sets to inform all parameters in DETECT.

4.6 Non-steady state model of soil CO$_2$ transport and production

An important contribution of this study was the development of a non-steady state (NSS) model of soil CO$_2$ transport and production (the DETECT model version 1.0), which is particularly useful for systems that may frequently experience NSS conditions. Other comparable NSS models exist (e.g., Šimůnek and Suarez, 1993; Fang and Moncrieff, 1999; Hui and Luo, 2004), but they generally treat the production (source) terms—root/rhizosphere respiration and microbial decomposition of soil organic matter—simplistically, and accompanying model code is not available. Our DETECT v1.0 model includes more detailed submodels for the production
terms, inspired by recent studies (E.g. Lloyd and Taylor, 1994; Pendall et al., 2003; Davidson et al., 2012; Todd-Brown et al., 2012; Carrillo et al., 2014a); in contrast to these studies, which essentially described models for “bulk” soil, we applied the CO$_2$ production models to every depth increment. Additionally, we have provided model code, implemented in Matlab (see Code Availability section), with the goal of making the DETECT model, and ability to accommodate NSS conditions, more accessible to potential users.

Future versions of DETECT could include other characteristics of soil CO$_2$ production and transport not included in v1.0. These include: (1) a transport process that simulates the physical displacement of CO$_2$ in the soil following a precipitation event; (2) alternative options for some of the functions used, for example there are a number of ways of estimating soluble soil C from soil organic C and soil water content (equation 7); (3) estimation of the parameters and their associated uncertainties using formal methods (e.g. MCMC) that rely on measurements of C stocks and C fluxes; (4) quantification of the uncertainty of the model outputs (soil CO$_2$ concentration, soil respiration) by propagation of uncertainty from the parameters; (5) coupling DETECT with a dynamic soil C model in order for the C$_{SOM}$ pools to be dynamic rather than prescribed independently of DETECT.

5. Conclusions

Determining the conditions under which steady-state (SS) assumptions are appropriate for modeling soil CO$_2$ production, transport, and efflux is crucial for accurately modeling the contribution of soils to the carbon cycle. We found that soil texture exerted the greatest control over whether SS assumptions are appropriate. When the soil at a site is coarse (60% or more sand), SS assumptions appeared to be appropriate, and one could apply a simpler, more computationally efficient SS model, such as SS-DETECT (see also Amundson et al., 1998). As
the soil texture becomes increasingly finer, SS assumptions start to break down, especially following large precipitation events that can greatly impact soil water content and associated soil air-filled porosity, thus affecting CO₂ diffusivity. Under such conditions, the more complex and computationally demanding NSS model (DETECT) is preferred. We found that precipitation regime characteristics and/or the inclusion of antecedent soil moisture and temperature conditions had little singular effect on whether SS or NSS assumptions were appropriate. However, while these factors do not directly impact SS versus NSS behavior, they were found to be important for accurately modeling the soil carbon cycle because they notably impacted the magnitude of the soil CO₂ efflux.

Code availability

All of the Matlab script files for running the DETECT model can be accessed via http://doi.org/10.5281/zenodo.927501. These Matlab script files are set up so that the model runs at the PHACE field site. The above weblink also provides a user manual which gives instructions for running DETECT at either the PHACE site or at a user specified field site. We also provide Matlab script files for creating a time series of predicted versus observed soil respiration (figure 1) and a time series of predicted versus observed soil CO₂ (figure 5). These can be found via http://doi.org/10.5281/zenodo.927313. Following publication, these Matlab files and the data files (see next section) will be available to download from the Ogle lab website via http://jan.ucc.nau.edu/ogle-lab/.

Data availability

Measurement data made at the PHACE field site, which are required as inputs for the DETECT model, are available via http://doi.org/10.5281/zenodo.926064.
Acknowledgements

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Figures and Tables

Figure 1. Graphical representation of (A) the required inputs to the DETECT model and the associated scenarios implemented in this study, (B) the components of the DETECT model at a particular time $t$, indicating depth-dependent production, $\text{CO}_2$ concentrations, and $\text{CO}_2$ fluxes, and (C) example model outputs, such as temporally and spatially varying $\text{CO}_2$ diffusivity and $\text{CO}_2$ production, and temporally varying bulk $\text{CO}_2$ fluxes.
**Figure 2** Time-series of daily surface soil CO$_2$ fluxes ($R_{\text{soil}}$) predicted by the non-steady-state (DETECT) and steady-state (SS-DETECT) models over the growing season (1$^{\text{st}}$ April – 30$^{\text{th}}$ September), based on the control scenarios (a) without ($Ctrl$) and (b) with ($Ctrl$-$\text{ant}$) antecedent effects (see Table 2). Only $R_{\text{soil}}$ is simulated using the SS-DETECT model, whereas $R_{\text{soil}}$ and its root and microbial contributions are simulated using the DETECT model. The predicted fluxes are overlaid with observed ecosystem respiration ($R_{\text{eco}}$; $R_{\text{soil}}$ + aboveground plant respiration) and microbial respiration ($R_{\text{m}}$; based on plots where vegetation was removed).

Commented [E43]: RC2, specific comment #22. Legend typo is now fixed (i.e. root $R$ = green line, microbial $R$ = blue line).
Figure 3  Time-series of daily surface soil respiration ($R_{soil}$) predicted from the non-steady-state (NSS) DETECT model (red solid lines) and the steady-state (SS-DETECT) model (grey dashed lines), for different soil texture scenarios. The first three scenarios are the same as the control (Ctrl), except they assume a different soil texture: (a) more sandy soil, (b) more silty soil, or (c) more clayey soil. Panels (d), (e), and (f) show the $R_{soil}$ predictions from the same soil texture scenarios as in (a)-(c), but also including antecedent effects of soil moisture and temperature. See Table 2 for descriptions of each scenario. $R_{soil}$ predictions are overlaid with daily precipitation.
Figure 4 Differences of total growing season (April-September) soil respiration ($R_{soil}$) as predicted by the non-steady-state (DETECT) and steady-state (SS-DETECT) models, for different pairs of scenarios. Comparisons are grouped such that they quantify the effects of (a) soil texture without antecedent effects, (b) soil texture with antecedent effects, (c) precipitation without antecedent effects, (d) precipitation with antecedent effects, and (e) antecedent effects. See Table 2 for descriptions of each scenario.
Figure 5 Time-series of predicted versus observed soil CO$_2$ concentrations at 3 cm depth, 10 cm depth, and 20 cm depth, where the predictions are based on the non-steady-state (NSS) DETECT model. Predicted [CO$_2$] is shown for the daily time-scale for the control scenarios (a) without (Ctrl) and (b) with (Ctrl-ant) antecedent effects, and for (c) the subdaily (every 6 hours) time scale for the Ctrl-ant scenario. Units are in parts per million (ppm).
Figure 6 Time series of how the modelled diffusivity of CO$_2$ ($D_{gs}$) at three different depths (5, 25, and 50 cm) varies between a predominantly sandy soil (solid line) and a predominantly clay soil (dashed line). Predictions are from the non-steady state (DETECT) model for the Ctrl (60% sand) and STLCl (60% clay) scenarios; see Table 2 for a description of the scenarios.
Table 1 Summary of scalar parameters used in the non-steady-state (DETECT) model, arranged into four groups: parameters unique to the microbial respiration submodel for $S_M(z,t)$ (group 1); parameters unique to the root respiration submodel for $S_R(z,t)$ (group 2); parameters that are shared between the $S_M(z,t)$ and $S_R(z,t)$ submodels (group 3); parameters used to calculate soil CO$_2$ diffusivity, $D_{liq}$ (group 4). See § 2.4.5 for details about how the parameters were estimated.

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Description</th>
<th>Value</th>
<th>Units</th>
<th>Eqn(s).</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Group 1 – root submodel parameters</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$R^*$</td>
<td>Total root biomass C in a 1 m deep by 1 cm$^2$ soil column</td>
<td>111.5</td>
<td>mg C cm$^{-2}$</td>
<td>3</td>
</tr>
<tr>
<td>$R_{RBase}$</td>
<td>Root mass-base respiration rate at 10 °C and mean environmental conditions</td>
<td>$6 \times 10^{-5}$</td>
<td>mg C cm$^{-3}$ hr$^{-1}$</td>
<td>3</td>
</tr>
<tr>
<td>$\alpha_1(R)$</td>
<td>The effect of soil water content ($\theta$) on root respiration</td>
<td>11.65</td>
<td>unitless</td>
<td>3, 4a</td>
</tr>
<tr>
<td>$\alpha_2(R)$</td>
<td>The effect of antecedent $\theta$ ($\theta_{Mant}$) on root respiration</td>
<td>20.7</td>
<td>unitless</td>
<td>3, 4b</td>
</tr>
<tr>
<td>$\alpha_3(R)$</td>
<td>The interactive effect of $\theta$ and $\theta_{Mant}$ on root respiration</td>
<td>-164.2</td>
<td>unitless</td>
<td>3, 4c</td>
</tr>
<tr>
<td><strong>Group 2 – microbial submodel parameters</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>$S^*$</td>
<td>Total soil organic C in a 1 meter deep by 1 cm$^2$ soil column</td>
<td>711.6</td>
<td>mg C cm$^{-2}$</td>
<td>5</td>
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<tr>
<td>$M^*$</td>
<td>Total microbial biomass C in a 1 meter deep by 1 cm$^2$ soil column</td>
<td>12.3</td>
<td>mg C cm$^{-2}$</td>
<td>5</td>
</tr>
<tr>
<td>$V_{Base}$</td>
<td>Value of $V_{max}$ at 10 °C and mean environmental conditions</td>
<td>0.0015</td>
<td>mg C cm$^{-3}$ hr$^{-1}$</td>
<td>5, 6</td>
</tr>
<tr>
<td>$\alpha_1(M)$</td>
<td>The effect of $\theta$ on microbial respiration</td>
<td>14.05</td>
<td>unitless</td>
<td>5, 6</td>
</tr>
<tr>
<td>$\alpha_2(M)$</td>
<td>The effect of antecedent $\theta$ ($\theta_{Mant}$) on microbial respiration</td>
<td>11.05</td>
<td>unitless</td>
<td>5, 6</td>
</tr>
<tr>
<td>$\alpha_3(M)$</td>
<td>The interactive effect of $\theta$ and $\theta_{Mant}$ on microbial respiration</td>
<td>-87.6</td>
<td>unitless</td>
<td>5, 6</td>
</tr>
<tr>
<td>$K_m$</td>
<td>Michaelis-Menton half saturation constant</td>
<td>$10^{-5}$</td>
<td>mg C cm$^{-3}$ hr$^{-1}$</td>
<td>5</td>
</tr>
<tr>
<td>$CUE$</td>
<td>Microbial carbon-use efficiency</td>
<td>0.8</td>
<td>mg C / mg C</td>
<td>5</td>
</tr>
<tr>
<td>$p$</td>
<td>Fraction of soil organic C that is soluble</td>
<td>0.004</td>
<td></td>
<td>7</td>
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<tr>
<td>$D_{liq}$</td>
<td>Diffusivity of soil C substrate in liquid</td>
<td>3.17</td>
<td>unitless</td>
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<tr>
<td><strong>Group 3 – shared parameters between root / microbial submodels</strong></td>
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<tr>
<td>$E_o^*$</td>
<td>Temperature sensitivity parameter, somewhat analogous to an energy of activation</td>
<td>324.6</td>
<td>Kelvin</td>
<td>4e</td>
</tr>
<tr>
<td>$T_o$</td>
<td>Temperature sensitivity-related parameter</td>
<td>227.5</td>
<td>Kelvin</td>
<td>4e</td>
</tr>
<tr>
<td>$\alpha_4$</td>
<td>The effect of antecedent soil temperature ($T_{Mant}$) on root and microbial respiration</td>
<td>-4.7</td>
<td>unitless</td>
<td>4e</td>
</tr>
<tr>
<td><strong>Group 4 – soil CO$_2$ diffusivity submodel parameters</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>$\alpha_3(R)$</td>
<td>Absolute value of the slope of the line relating log($\Psi$) versus log($\theta$)</td>
<td>4.547</td>
<td>unitless</td>
<td>2</td>
</tr>
<tr>
<td>$BD$</td>
<td>Soil bulk density</td>
<td>1.12</td>
<td>g cm$^{-3}$</td>
<td>2</td>
</tr>
<tr>
<td>$\phi_{100}$</td>
<td>Air-filled porosity at soil water potential of -100 cm H$_2$O (~10 kPa)</td>
<td>18.16</td>
<td>%</td>
<td>2</td>
</tr>
<tr>
<td>$PD$</td>
<td>Particle density</td>
<td></td>
<td></td>
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</table>
Table 2 Summary of quantities in the non-steady-state (DETECT) model that vary by depth only (z), or by depth (z) and time (t). Those in group 1 represent input variables (derived prior to the running of the DETECT model), while group 2 contains the modeled quantities (used as part of the operation of the DETECT model). Equation S1 can be found in Appendix S2 in the supplementary information.

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Description</th>
<th>Units</th>
<th>Eqn(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>( f_R(z) )</td>
<td>A function describing the distribution by depth of root carbon.</td>
<td>unitless</td>
<td>S1</td>
</tr>
<tr>
<td>( C_R(z, t) )</td>
<td>The amount of root carbon.</td>
<td>mg C cm(^{-1}) hr(^{-1})</td>
<td>3, S1</td>
</tr>
<tr>
<td>( f_S(z) )</td>
<td>A function describing the distribution by depth of carbon from soil organic matter (SOM)</td>
<td>unitless</td>
<td>S1</td>
</tr>
<tr>
<td>( C_{SOM}(z) )</td>
<td>The amount of carbon from SOM.</td>
<td>mg C cm(^{-1}) hr(^{-1})</td>
<td>7, S1</td>
</tr>
<tr>
<td>( f_M(z) )</td>
<td>A function describing the distribution by depth of microbial carbon</td>
<td>unitless</td>
<td>S1</td>
</tr>
<tr>
<td>( C_{MIC}(z) )</td>
<td>The amount of microbial carbon.</td>
<td>mg C cm(^{-1}) hr(^{-1})</td>
<td>3, S1</td>
</tr>
<tr>
<td>( \theta(z, t) )</td>
<td>Soil water content</td>
<td>m(^3) m(^{-3})</td>
<td>3, 6, 7</td>
</tr>
<tr>
<td>( \theta_{ant}(z, t) )</td>
<td>Antecedent soil water content (used in ( S_R ) function) calculated as a weighted average of soil water content from the previous 4 days. The weights are ( w=(0.75,0.25,0,0) ).</td>
<td>m(^3) m(^{-3})</td>
<td>3</td>
</tr>
<tr>
<td>( \theta_{ant}(z, t) )</td>
<td>Antecedent soil water content (used in ( S_M ) function) calculated as a weighted average of soil water content from the previous 4 days. The weights are ( w=(0.2,0.6,0.2,0) ).</td>
<td>m(^3) m(^{-3})</td>
<td>6</td>
</tr>
<tr>
<td>( T_S(z, t) )</td>
<td>Soil temperature</td>
<td>Kelvin</td>
<td>3, 6</td>
</tr>
<tr>
<td>( T_{ant}(z, t) )</td>
<td>Antecedent soil temperature calculated as a weighted average of soil temperature from the previous 4 weeks. The weights are ( w=(0.25,0.25,0.25,0.25) ).</td>
<td>Kelvin</td>
<td>3, 6</td>
</tr>
<tr>
<td>( c(z, t) )</td>
<td>Total soil CO(_2).</td>
<td>mg CO(_2) m(^{-3})</td>
<td>1</td>
</tr>
<tr>
<td>( c_R(z, t) )</td>
<td>Soil CO(_2) derived from root sources.</td>
<td>mg CO(_2) m(^{-3})</td>
<td>1</td>
</tr>
<tr>
<td>( S_R(z, t) )</td>
<td>Source term describing the production of soil CO(_2) from root respiration.</td>
<td>mg CO(_2) m(^{-3})</td>
<td>1</td>
</tr>
<tr>
<td>( c_m(z, t) )</td>
<td>Soil CO(_2) derived from microbial sources.</td>
<td>mg CO(_2) m(^{-3})</td>
<td>1</td>
</tr>
<tr>
<td>( S_m(z, t) )</td>
<td>Source term describing the production of soil CO(_2) from microbial respiration.</td>
<td>mg CO(_2) m(^{-3})</td>
<td>1</td>
</tr>
<tr>
<td>( D_{gs}(z, t) )</td>
<td>Diffusivity of soil CO(_2).</td>
<td>m(^2) s(^{-1})</td>
<td>1, 2</td>
</tr>
<tr>
<td>( \phi(z, t) )</td>
<td>Air-filled soil porosity.</td>
<td>m(^3) m(^{-3})</td>
<td>1, 2</td>
</tr>
<tr>
<td>( C_{SOL}(z, t) )</td>
<td>The amount of soluble carbon from SOM.</td>
<td>mg C cm(^{-1}) hr(^{-1})</td>
<td>5, 7</td>
</tr>
<tr>
<td>( V_{max}(z, t) )</td>
<td>Maximum potential decomposition rate (microbial carbon).</td>
<td>mg C cm(^{-1}) hr(^{-1})</td>
<td>6</td>
</tr>
<tr>
<td>( E_o(z, t) )</td>
<td>Analogous to energy of activation.</td>
<td>Kelvin</td>
<td>4c</td>
</tr>
</tbody>
</table>


1 Table 3 The scenario code, description, and summary of results associated with each model scenario; the 14 scenarios below were
2 applied to both the DETECT and SS-DETECT models. The scenarios involved a non-factorial combination of different soil texture,
3 precipitation regimes, and inclusion/exclusion of antecedent effects on the root and microbial CO₂ production rates.

<table>
<thead>
<tr>
<th>Scenario</th>
<th>Description</th>
<th>Primary result(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ctrl (control)</td>
<td>Uses soil texture (sandy clay loam: 60% sand, 20% clay) and precipitation</td>
<td>$R_{\text{soil}}$ was very similar under SS and NSS soil CO₂ assumptions.</td>
</tr>
<tr>
<td></td>
<td>(for 2008) data from the PHACE site; CO₂ production only responds to</td>
<td></td>
</tr>
<tr>
<td></td>
<td>concurrent environmental conditions.</td>
<td></td>
</tr>
<tr>
<td>Soil texture scenarios</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ST-Sa</td>
<td>Same as Ctrl, but the soil texture is set to sandy loam (80% sand, 10% clay).</td>
<td>For ST-Cl, $R_{\text{soil}}$ was greater in magnitude and more different under SS vs NSS conditions, due to NSS conditions producing greater $R_{\text{soil}}$ after a major precipitation event. The results are similar, but muted, for the ST-Si scenario.</td>
</tr>
<tr>
<td>ST-Si</td>
<td>Same as Ctrl, but the soil texture is set to silt loam (20% sand, 20% clay).</td>
<td></td>
</tr>
<tr>
<td>ST-Cl</td>
<td>Same as Ctrl, but the soil texture is set to clay (20% sand, 60% clay).</td>
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<tr>
<td>Precipitation scenarios</td>
<td></td>
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<tr>
<td>P-E</td>
<td>Same as Ctrl, but daily precipitation was shifted to occur one month earlier.</td>
<td>Varying the timing or magnitude of precipitation pulses had little effect on the magnitude of $R_{\text{soil}}$ or on the difference between SS and NSS predictions of $R_{\text{soil}}$.</td>
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<tr>
<td>P-L</td>
<td>Same as Ctrl, but daily precipitation was shifted to occur one month later.</td>
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</tr>
<tr>
<td>P-FM</td>
<td>Same as Ctrl, but daily precipitation was based on data from 2009, which is characterized by more frequent, smaller events.</td>
<td></td>
</tr>
</tbody>
</table>

**Scenarios that incorporate antecedent effects on CO₂ production rates**

<table>
<thead>
<tr>
<th>Scenario</th>
<th>Description</th>
<th>Primary result(s)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ctrl-ant</td>
<td>All scenarios parallel those described above, except both current and</td>
<td>$R_{\text{soil}}$ was generally greater in magnitude under both SS and NSS conditions, especially for ST-Si-ant and ST-Cl-ant (relative to ST-Si and ST-Cl).</td>
</tr>
<tr>
<td>ST-Sa-ant</td>
<td>antecedent conditions (past soil water and past soil temperature) are used in the calculation of the source terms (i.e., root and microbial CO₂ production rates).</td>
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<tr>
<td>ST-Si-ant</td>
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<tr>
<td>ST-Cl-ant</td>
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<tr>
<td>P-E-ant</td>
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<tr>
<td>P-L-ant</td>
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<tr>
<td>P-FM-ant</td>
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</tbody>
</table>